Transcutaneous CO$_2$ application accelerates fracture repair in streptozotocin-induced type I diabetic rats

Takahiro Oda 1, Takahiro Niikura 1, Tomoaki Fukui 1, Keisuke Oe 1, Yu Kuroiwa 1, Yohei Kumabe 1, Kenichi Sawauchi 1, Ryo Yoshikawa 1, Yutaka Mifune 1, Shinya Hayashi 1, Tomoyuki Matsumoto 1, Takehiko Matsushita 1, Teruya Kawamoto 1, Yoshitada Sakai 2, Toshihiro Akisue 3, Ryosuke Kuroda 1

INTRODUCTION

Diabetes mellitus (DM) is a factor that can negatively affect bone fracture healing. Delayed fracture healing and non-union are more frequent in patients with diabetes, and bony union in such patients often requires a longer period. In studies using closed femoral shaft fracture models in DM rats, several authors have demonstrated that delayed fracture healing occurs because of inhibition of endochondral ossification and callus formation. In the clinical setting, the number of patients with diabetes is predicted to increase sharply to 693 million worldwide by 2045. Therefore, establishing the causes of delayed fracture healing and developing novel approaches to treat it in patients with diabetes are vital.

We previously designed an easy, non-invasive, topical system for transcutaneous application of CO$_2$ gas using hydrogel as a novel approach to promote bone fracture healing and continued our investigation for clinical application in fracture treatment. We demonstrated that transcutaneous CO$_2$...
application facilitates O₂ dissociation from hemoglobin via the Bohr effect, thereby promoting oxygenation in the tissues and enhancing local blood flow. Our previous study using a rat femoral fracture model showed that transcutaneous application of CO₂ accelerated fracture repair by promoting angiogenesis, blood flow, and endochondral ossification at the fracture site. Although the detailed molecular or cellular mechanisms are still unclear, transcutaneous CO₂ application might increase vascular endothelial growth factor (VEGF) expression around the fracture site, accelerating the fracture healing process via enhanced angiogenesis.

Therefore, using a diabetic rat fracture model, we aimed to determine whether transcutaneous CO₂ application would promote bone fracture healing in DM by accelerating endochondral ossification and angiogenesis.

RESEARCH DESIGN AND METHODS

Animals

Altogether, 90 female Sprague Dawley rats (SLC Japan, Shizuoka, Japan) with a mean weight (±SD) of 228.4 g (±5.6 g) and 12 weeks old were used in this study. DM rats were created by a single intraperitoneal administration of 40 mg/kg streptozotocin (STZ) (Sigma-Aldrich, St. Louis, MO); the experimental model had type 1 diabetes. Fractures were induced 2 weeks after STZ administration. Blood glucose levels were measured at the time of fracture creation and sacrifice. Animals with blood glucose levels <300 mg/dL were not recognized as DM rats and were excluded from the study. In this study, we determined the sample size with reference to our previous study.

Surgical procedure

A standard stabilized closed femoral shaft fracture was induced according to previously reported methods. Briefly, retrograde insertion of a 1.25-mm-diameter K-wire into the right femoral intramedullary canal was performed, followed by induction of a closed transverse femoral shaft fracture using a three-point bending apparatus and a drop weight. After creating the fracture, the patellar aponeurosis and the skin were sutured with nylon. Preoperatively, we administered medetomidine (0.15 mg/kg), midazolam (2 mg/kg), and butorphanol (2.5 mg/kg) intraperitoneally for anesthesia and sedation. Postoperatively, we administered benzylpenicillin potassium (200 000 units/kg) intraperitoneally as an antibiotic agent. Unprotected weight bearing was allowed postoperatively. The rats were sacrificed using pentobarbital overdose before the assessments.

Transcutaneous application of CO₂

The animals were randomly divided into two groups: the CO₂ group and the control group (n=45 in each group). Transcutaneous CO₂ was applied to the fractured lower limbs of the rats previously reported.7 Briefly, after induction of sedation with a minimum dose of isoflurane, the fractured limb was shaved, and hydrogel (NeoChemir, Kobe, Japan), which enhances CO₂ absorption, was applied. The hydrogel (pH 5.5) consisted of carbomer, glycerin, sodium hydroxide, sodium alginate, sodium dihydrogen phosphate, methylparaben, and deionized water. Both limbs were sealed with a polyethylene bag, which was filled with 100% CO₂ for 20 min (online supplemental figure S1). This treatment was performed 5 days a week. The control group received sham treatment, where CO₂ was replaced with air.

Radiographic assessment of fracture repair

The anesthetized rats were fixed in the supine position with the limbs fully extended, and radiographs of the fractured limbs were obtained (n=10 in each group at 1, 2, and 3 weeks after fracture; n=15 in each group at 4 weeks after fracture). Fracture union was defined as the presence of bridging callus formation in at least three of the four cortices on the anteroposterior and lateral views.

Histological assessment for fracture sites

Histological assessment was performed with Safranin-O staining at 1, 2, 3, and 4 weeks after fracture (n=5 in each group). Harvested femurs were fixed in 4% paraformaldehyde at room temperature for 24 hours. Subsequently, the femurs were decalcified at room temperature with a decalifying solution (10% formic acid and 10% formalin; 1:1 ratio) and embedded in paraffin wax. Finally, the femurs were processed to obtain 6-μm sagittal sections using a microtome. The sections were deparaffinized in xylene, dehydrated in a graded alcohol series, stained with Safranin-O/fast green, and detailed histological structures and cartilage areas were visualized using a light microscope. To assess the progression of endochondral ossification, the total cartilage area was calculated using National Institutes of Health (NIH) ImageJ software. The degree of fracture repair was assessed on a five-point scale (grade 0: Pseudoarthrosis formation, grade 1: Incomplete cartilaginous union, grade 2: Complete cartilaginous union, grade 3: Incomplete bony union, grade 4: Complete bony union) according to Allen’s grading system.

To evaluate the intrarater interclass correlation coefficient (ICC) and the inter-rater ICC reproducibility, determinations of bone union and scoring of degree of fracture repair by Allen’s grading system were performed twice by five surgeons blindly.

Assessment of angiogenesis

At weeks 1, 2, 3, and 4 after fracture, angiogenesis was evaluated (n=5 in each group). To evaluate cross-sectional capillary density, immunofluorescence staining of endothelial cells was performed using fluorescein-labeled isocitrate B4 (Vector Laboratories, Burlingame, California, USA). Nuclear staining was performed using 4′,6-diamidino-2-phenylindole (DAPI) solution (Nacalai Tesque, Kyoto, Japan). Capillaries were morphometrically examined under a fluorescent microscope. The capillaries in five randomly selected fields in the granulation tissue around the fracture site were counted, and the means were calculated.
Immunohistochemical analysis

Immunohistochemical assessment was performed at 2 weeks and 3 weeks after the fracture (n=5 in each group). We assessed Ki67 expression of chondrocyte within bony callus because it is one of the best-known proliferation markers.\(^{11,12}\) We also evaluated cathepsin K expression callus because it is one of the best-known osteoclast markers.\(^ {13,14}\) The sections were incubated overnight at 4°C with anti-Ki67 antibody (1:50 dilution, NB500-170, Novus Biologicals, Centennial, Colorado, USA) or anticathepsin K antibody (1:50 dilution, ab19027, Abcam, Cambridge, Massachusetts, USA) and subsequently treated with peroxidase-labeled antimouse immunoglobulin (Histofine Simple Stain MAX PO (R), Nichirei Bioscience, Tokyo, Japan) at room temperature for 60 min. The signal was developed as a brown reaction product using the peroxidase substrate 3,3′-diaminobenzidine (Histofine Simple Stain 3,3′-Diaminobenzidine (DAB) Solution, Nichirei Bioscience). The sections were counterstained with hematoxylin and examined with a BZ-X700 confocal microscope (Keyence Corporation, Osaka, Japan). Immunopositive cells were counted in four random fields under a high-power field.

All morphometric studies of immunofluorescence and immunohistochemical staining were performed by two blinded orthopedic surgeons.

Gene expression assessment and RNA extraction

At weeks 1, 2, 3, and 4 after fracture, the expression of specific genes was measured in five animals in each group by real-time PCR. Newly generated callus tissues around the fracture site were harvested. Tissues were homogenized using TRIzol (Invitrogen, Carlsbad, California, USA) with a T18 ULTRA-TURRAX homogenizer (IKA Werke, Staufen, Germany) immediately after harvesting. Total cellular RNA was extracted from the harvested tissues using the acid guanidinium thiocyanate-phenol-chloroform method and purified using RNeasy Mini Kit (Qiagen, Valencia, California, USA); subsequently, on-column digestion was performed using RNase-free DNase Kit (Qiagen) to prevent contamination with genomic DNA.

Real-time PCR

RNA samples were reverse-transcribed to synthesize single-stranded complementary DNA (cDNA) samples using a high-capacity cDNA reverse transcriptional kit (Applied Biosystems, Foster City, California, USA) according to the manufacturer’s instructions. Real-time PCR was performed in duplicate on the cDNA with ABI PRISM 7700 Sequence Detection System and SYBR Green reagent (Applied Biosystems). All primer sequences are shown in online supplemental table S1.

We examined gene expression levels of collagen II, collagen X, and matrix metalloproteinase-13 (MMP-13) to evaluate chondrogenic differentiation; runt-related transcription factor 2 (Runx2) and osterix was used to evaluate osteogenic differentiation; and VEGF was used to evaluate angiogenesis. We evaluated gene expression levels of tumor necrosis factor-α (TNF-α), macrophage colony-stimulating factor (M-CSF), receptor activator of NFκB (nuclear factor-kappa B) ligand (RANKL), and a disintegrin and metalloproteinase with thrombospondin motifs 4 (ADAMTS 4), all of which have been reported to inhibit fracture healing by being overexpressed in DM.\(^ {15,16}\) We also examined endothelial nitric oxide synthase (eNOS), which contributes to vasodilation function,\(^ {17}\) and thrombospondin-1 (TSP-1), which is a potent endogenous inhibitor of angiogenesis.\(^ {18}\) The expression levels of each gene were first normalized with respect to that of GAPDH (glyceraldehyde-3-phosphatdehydrogenase), which served as an internal control. All results are presented as the fold change relative to one sample of the control group at week 1 (ΔΔCT method; Applied Biosystems).\(^ {19}\)

Biomechanical assessment for fracture repair

A standardized three-point bending test was performed using a load torsion and bending tester at week 4 after fracture (n=5 in each group). The bending force was applied with the crosshead at a speed of 2mm/min until rupture occurred. Ultimate stress (N), extrinsic stiffness (N/mm), and failure energy (N·mm) were measured.

Statistical analysis

Fisher’s exact test was used for radiographic assessment. Histological (degree of fracture repair), fluorescent immunohistochemical, and biomechanical assessments, as well as genetic evaluation with real-time PCR, were performed using the Mann-Whitney U test. The Kruskal-Wallis test with the Bonferroni-corrected post hoc Mann-Whitney U test was used for histological (cartilage area size comparison) and immunohistochemical assessments. P values <0.05 were considered statistically significant. Columns and error bars indicate means and standard errors, respectively. All statistical analyses were carried out using BellCurve for Excel V.3.0 (Social Survey Research Information, Tokyo, Japan).

RESULTS

Radiographic assessment of fracture repair

Representative radiographs of both groups at each time point are shown in figure 1. At week 1, periosteal callus formation was not observed in either the CO2 or control groups (figure 1A,E). At week 2, bony callus became visible in the CO2 group but not in the control group (figure 1B,F; white arrows). Bony callus was observed at week 3 in the control group; however, the callus size was not sufficient to form bridging callus even at week 4 (figure 1C,D). At week 3, fracture union with bridging callus formation was achieved in 50% (5/10) of the animals in the CO2 group and 20% (2/10) in the control group. At week 4, fracture union was achieved in 80% (12/15) in the CO2 group and 20% (3/15) in the control group. The fracture union rate at week four was significantly higher in the CO2 group than in the control group (p<0.05; figure 1).
Biomechanical assessment for fracture repair

All three evaluation items for biomechanical assessment (ie, ultimate stress, extrinsic stiffness and failure energy) were significantly higher in the CO2 group than in the control group (p<0.05; figure 5).

DISCUSSION

DM can delay fracture repair and cause failure of fracture union. Our study showed that transcutaneous CO2 application accelerated fracture repair even in cases with DM.

DM inhibits the bone fracture repair process by impairing cartilage differentiation in the early phase, delaying endochondral ossification, and reducing biomechanical properties of fracture callus.20 Nevertheless, the detailed mechanisms by which DM inhibits bone fracture repair remain unclear. To elucidate the molecular mechanisms, various studies using rats with type I diabetes have been conducted.20 21 Several authors reported that...
Genetics/Genomes/Proteomics/Metabolomics

Diabetic rats require a longer time to achieve fracture union than do healthy rats. Ogasawara et al demonstrated that bone union is achieved at 6 weeks in DM rat femoral fracture model, similar to the model in our study. Moreover, in our study, at 4 weeks after fracture, bone union was achieved in 80% of DM rats in the CO2 group and in only 20% of the rats in the control group, suggesting that transcutaneous CO2 application accelerated fracture repair. Histologically, maturation and hypertrophy of chondrocytes are suppressed and remarkably small cartilage callus is formed in DM rats than in healthy rats. In this study, although it remains unknown whether the cartilage area of DM rats in the control group was smaller than that of healthy rats, the cartilage area was significantly larger in the CO2 group than in the control group at weeks 1 and 2, suggesting that chondrocyte differentiation is promoted by transcutaneous CO2 application. This is consistent with the results of immunohistochemical staining for Ki67, in which the percentage of proliferating cells was significantly higher in the CO2 group throughout the fracture repair period. These two histological findings suggest that the fracture repair process is accelerated through the promotion of endochondral ossification by transcutaneous CO2 application. In the biomechanical assessment, all evaluation items showed significantly greater values in the CO2 group, and the improvement in the reduction of biomechanical properties due to DM could be associated with the transcutaneous CO2 application.

Angiogenesis and blood flow at the fracture site are essential factors for fracture repair. In the inflammatory phase of fracture healing, early blood vessel formation supports the invasion of inflammatory cells into the fracture site, and the inflammatory cells release various cytokines that are necessary for fracture repair. Endochondral ossification, which is one of the most important processes in fracture healing, starts during the reparative phase. During this process, avascular cartilaginous tissue is invaded by blood vessels and is transformed into vascular osseous tissue. Therefore, active angiogenesis around the fracture site is indispensable in this phase. Furthermore, sufficient blood flow is essential to
supply calcium and phosphate that is needed for callus mineralization following endochondral ossification. In a rat experimental model, inhibition of angiogenesis suppressed fracture repair. We previously reported that the capillary density around the fracture site was significantly lower in diabetic rats than in healthy rats. Inhibition of angiogenesis around the fracture site could be a factor of delayed fracture repair in diabetic animals. Various growth factors have been reported to be involved in the fracture repair process. VEGF is one of the most important genes associated with angiogenesis, and its expression is essential for normal angiogenesis, callus formation, and mineralization in fracture repair. Lim et al. reported that diabetes-induced TNF-α dysregulation has negative effects on angiogenesis during fracture repair by reducing vessel formation and VEGF expression. In the present study, we showed that gene expression levels of VEGF in newly generated callus tissue were significantly higher in the CO₂ group at all time points. Fluorescent immunostaining with isoelectin B4 revealed that angiogenesis around the fracture site was promoted in the CO₂ group. These results suggest that transcutaneous CO₂ application reverses the reduced gene expression levels of VEGF around the fracture site and results in sufficient angiogenesis for fracture repair. We previously reported that transcutaneous CO₂ application increased local oxygen partial pressure via the Bohr effect, resulting in a lower local pH. Acidosis is known to increase gene expression levels of VEGF. Taken together, these findings suggest that transcutaneous CO₂ application reduces the pH and increases expression levels of VEGF.

Transcutaneous CO₂ application increases local blood flow and therefore has been applied clinically to improve ischemic limb symptoms. Increased blood flow elevates wall shear stress in blood vessels, in turn inducing eNOS upregulation. eNOS is involved in vasodilation and in osteoblast maturation and activity. Other studies found that the activity and expression of eNOS were reduced in diabetes and hypoxia, and some reports showed that VEGF upregulated eNOS expression. In our study, gene expression levels of eNOS were significantly higher in the CO₂ group at weeks 2–4 even in DM rats. eNOS expression upregulated by the increased blood flow and the greater expression of VEGF possibly contributed to the promotion of fracture repair through vasodilation and maturation and activation of osteoblasts, as previously described. By contrast, gene expression levels of TSP-1, which suppresses VEGF bioavailability and activity as well as nitric oxide signaling, was significantly lower in the CO₂ group than in the control group at weeks 3 and 4. This result suggests that transcutaneous CO₂ application promotes fracture healing via downregulation of TSP-1, which inhibits angiogenesis.

Collagens II and X are involved in chondrocyte differentiation, and their gene expression is suppressed in diabetes, resulting in decreased callus size via inhibition...
Figure 4  Mean expression with SE of the 12 genes of interest in the control and CO₂ groups at weeks 1, 2, 3, and 4 after fracture, as measured by quantitative real-time PCR (n=5 in each group). Gene expression levels were normalized to GAPDH and are presented as fold change relative to a sample of the control group at week 1. ADAMTS 4, a disintegrin and metalloproteinase with thrombospondin motifs 4; eNOS, endothelial nitric oxide synthase; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; M-CSF, macrophage colony-stimulating factor; MMP-13, matrix metallopeptinase-13; RANKL, receptor activator of NF-κB ligand; Runx2, runt-related transcription factor 2; TNF-α, tumor necrosis factor-α; TSP-1, thrombospondin-1; VEGF, vascular endothelial growth factor.

Figure 5  Biomechanical assessment for fracture repair with three-point bending test at week 4 after fracture. Mean values with SE are shown (n=5 in each group).
of chondrocyte differentiation and in reduced mechanical properties.\(^4\) In the present study, gene expression levels of *collagens II* and *X* were significantly higher in the CO\(_2\) group than in the control group from the early stage of fracture healing to endochondral ossification. These results were consistent with the histological assessment finding that showed significantly larger cartilage formation in the CO\(_2\) group than in the control group at the early stage after fracture creation.

On the other hand, an elevation of the gene expression level of *collagen II* means that chondrocytes are in the proliferative phase, but prolonging this period may trigger delayed fracture healing. In this study, the gene expression level of *collagen II* seemed to peak at week 3 in the CO\(_2\) group, but the difference between weeks 2 and 3 was not statistically significant (Kruskal-Wallis test with the Bonferroni-corrected post hoc Mann-Whitney U test). We also performed immunohistochemical staining for Ki67, which is expressed in the nucleus of proliferating cells and is often used to determine if the cell is in a proliferative or resting phase.\(^1\)\(^-\)\(^2\) The percentage of chondrocytes with immunopositive staining for Ki67 was significantly higher in the CO\(_2\) group than in the control group at week 2; however, this difference disappeared at week 3. Additionally, in the CO\(_2\) group, the percentage of Ki67-positive chondrocytes dropped significantly from week 2 to week 3 (p<0.05; figure 4). These results indicate that before reaching week 3, the active proliferative phase of chondrocytes, with elevation of gene expression of collagen II, has finished, and the cartilage resorption phase has already begun. We believe this suggests that transcutaneous CO\(_2\) application promoted chondrocyte proliferation at an early time point after fracture.

MMP-13 plays an important role in the maturation of chondrocytes and the induction of angiogenesis. In MMP-13-deficient mice, fracture healing was significantly delayed because of inhibition of endochondral ossification.\(^4\)\(^1\) In the present study, gene expression levels of MMP-13 were significantly higher in the CO\(_2\) group at weeks 2 and 3, corresponding to the endochondral ossification period. This result was consistent with the histological finding that the cartilage area rapidly decreased from week 2 in the CO\(_2\) group because of the progression of endochondral ossification. These genetic and histological results of *collagen II*, *collagen X*, and MMP-13 suggest that transcutaneous CO\(_2\) application promotes chondrocyte differentiation and endochondral ossification, both of which are suppressed in diabetes, thereby resulting in improved fracture healing.

Runx2 and osterix are essential for osteoblast differentiation and both play vital roles in bone formation. Runx2 induces differentiation of mesenchymal progenitor cells to preosteoblasts, and osterix enables preosteoblast differentiation into immature osteoblasts.\(^4\)\(^2\) Runx2 and osterix are inhibited under hyperglycemic conditions, resulting in impaired bone formation and regeneration.\(^4\)\(^3\) In the present study, gene expression levels of both Runx2 and osterix were higher in the CO\(_2\) group at week 2 and later. Although the detailed mechanisms remain unclear, bone formation, which is typically impaired in diabetes, is possibly improved by transcutaneous CO\(_2\) application.

Overexpression of TNF-α and RANKL inhibits fracture healing in diabetes via overactivation of osteoclasts,\(^4\)\(^5\)\(^6\) nevertheless, no significant differences in their expression levels were noted between the groups in our study. In addition, immunohistochemical staining for cathepsin K performed at 2 weeks and 3 weeks after fracture, reported to be the period of excessive increase in the number of osteoclasts in diabetic fracture model animals,\(^4\)\(^5\)\(^6\) showed no statistically significant difference in osteoclast numbers between the CO\(_2\) group and the control group. These findings suggest that the promotion of fracture repair by transcutaneous CO\(_2\) application is probably not related to the osteoclast activity regulated by these genes.

M-CSF and ADAMTS 4 also inhibit fracture repair by the same mechanism as that of TNF-α and RANKL,\(^4\)\(^9\)\(^2\) however, their expression levels were significantly higher in the CO\(_2\) group than in the control group and peak at week 3 after fracture creation, corresponding to the time when active cartilage resorption in the fracture repair process in rats starts. We speculate that cartilage resorption was promoted in response to the increase in cartilage formation due to transcutaneous CO\(_2\) application.

Delayed fracture healing in patients with DM is a major issue to be solved. Prolonged treatment can lead to increased medical costs, reduce the level of activities of daily living among patients, and cause many complications, such as pneumonia. For this reason, various studies using DM animals have been conducted on promoting fracture healing under the condition of DM.\(^2\)\(^1\)\(^4\)\(^7\) However, the need to develop new tools to support fracture healing in DM continues, and the results of our study will be meaningful for fracture patients with DM. Transcutaneous application of CO\(_2\) is relatively safe and easy to perform without invasive surgery; however, clinical application still needs improvement. Ongoing research should be performed to improve the treatment’s convenience and safety in clinical settings.

Delayed wound healing and increased susceptibility to infection are also major problems in patients with DM. We believe that increasing blood flow through CO\(_2\) application may accelerate fracture healing and promote wound healing and support efficient local delivery of antibiotics to prevent and treat infection. Several authors have already reported the effects of application of CO\(_2\) on healing of wounds and ulcers.\(^8\)\(^9\) Our future studies will investigate the effectiveness of CO\(_2\) application as a support tool for treating wounds or preventing infection in patients with DM.

This study has some limitations. First, because it is difficult to reproduce in vivo conditions, in vitro evaluation could not be performed. Second, we included only rats with type I diabetes. A previous study reported that bone mineral density is decreased in type I diabetes but increased in type II diabetes.\(^5\)\(^0\) The detailed mechanisms of delayed fracture repair may differ between type I and
type II diabetes. The number of patients with type II diabetes is greater than those with type I diabetes; therefore, future studies should investigate animals with type II diabetes. Furthermore, whether similar results could be obtained with male rats and other animal species needs to be explored.

In conclusion, we found that transcutaneous application of CO₂ accelerated fracture healing by reversing the inhibition of cartilage formation, endochondral ossification, bone formation, and mechanical properties, all of which are reported to cause delayed fracture repair in DM. We believe this easy and less invasive treatment could be an effective strategy for patients with delayed fracture repair due to DM.

Acknowledgements The authors thank Ms K Tanaka, Ms M Nagata and Ms M Yasuda (Department of Orthopaedic Surgery, Kobe University Graduate School of Medicine) for their excellent technical assistance.

Contributors TO, YK, YK, and KS performed the surgeries and radiological examinations. TO performed the histological and genetic examinations. TO and TF analyzed all data, performed the statistical analysis, and were the major contributors in the writing of the manuscript. KO, KY, YM, SH, TM, TM, TK, YS, TA, FK, and TN conceived the study and participated in the study design and coordination. All authors read and approved the final manuscript. TN is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Funding This study is funded by Grant-in-Aid for Scientific Research (C) (JSPS KAKENHI Grant Number JP17K10966). The funding source had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests None declared.

Patient consent for publication Not required.

Ethics approval All procedures performed in the animals were in accordance with the ethical standards of the Animal Care and Use Committee of Kobe University Graduate School of Medicine (approval no. P170602-R1).

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement Data are available upon reasonable request. All data relevant to the study are included in the article or uploaded as supplemental information.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

REFERENCES