Multilevel clustering approach driven by continuous glucose monitoring data for further classification of type 2 diabetes

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ABSTRACT

Introduction Mining knowledge from continuous glucose monitoring (CGM) data to classify highly heterogeneous patients with type 2 diabetes according to their characteristics remains unaddressed. A refined clustering method that retrieves hidden information from CGM data could provide a viable method to identify patients with different degrees of dysglycemia and clinical phenotypes.

Research design and methods From Shanghai Jiao Tong University Affiliated Sixth People’s Hospital, we selected 908 patients with type 2 diabetes (18–83 years) who wore blinded CGM sensors (iPro2, Medtronic, California, USA). Participants were clustered based on CGM data during a 24-hour period by our method. The first level extracted the knowledge-based and statistics-based features to describe CGM signals from multiple perspectives. The Fisher score and variables cluster analysis were applied to fuse features into low dimensions at the second level. The third level divided subjects into subgroups with different clinical phenotypes. The four subgroups of patients were determined by clinical phenotypes.

Results Four subgroups of patients with type 2 diabetes with significantly different statistical features and clinical phenotypes were identified by our method. In particular, individuals in cluster 1 were characterized by the lowest glucose level factor and glucose fluctuation factor, and the highest negative glucose factor and C peptide index. By contrast, cluster 2 had the highest glucose level factor and the lowest C peptide index. Cluster 4 was characterized by the greatest degree of glucose fluctuation factor, was the most insulin-sensitive, and had the lowest insulin resistance. Cluster 3 ranked in the middle concerning the CGM-derived metrics and clinical phenotypes compared with those of the other three groups.

Conclusion A novel multilevel clustering approach for knowledge mining from CGM data in type 2 diabetes is presented. The results demonstrate that subgroups are adequately distinguished with notable statistical and clinical differences.

INTRODUCTION

Diabetes and its complications can substantially impair the human body. Its treatment has always been a long-standing dilemma worldwide.1 2 Type 2 diabetes not only comprises 90% of all diabetes, but also has considerable heterogeneity in genes and clinical phenotypes.3 4 It is generally believed that further classification of type 2 diabetes will provide more personalized and precise treatments for patients.3

Cluster analysis, a data-mining technique that helps reveal hidden structures, has been widely used in the medical field.5 6 Previous works using cluster analyses to distinguish patients with type 2 diabetes have largely concentrated on medical check-up data and...
They were diagnosed with type 2 diabetes according to Affiliated Sixth People’s Hospital from January 2018 to crinology and Metabolism, Shanghai Jiao Tong University Study population

METHODS

To the best of our knowledge, empirical research on classification of patients with type 2 diabetes from CGM data is limited. By dividing the complete CGM curve into segments, Hall and colleagues found three different fluctuation modes of CGM, and the frequencies of the three modes in individuals without diabetes, patients with pre-diabetes and patients with diabetes were finally found to be different. Kahkoska and colleagues used eight CGM features to identify new subgroups of type 1 diabetes. Three subgroups showed significant differences in glycated hemoglobin A1c (HbA1c). The problem of using CGM indices to stratify patients with type 2 diabetes among different categories needs to be discussed further.

As a result, examining patients with type 2 diabetes by CGM data and clustering these patients into specific groups may provide useful information for decision-making. In this study, we proposed a new data-driven method to further classify patients with type 2 diabetes into different subgroups from the original CGM data. We attempt to identify these distinct subgroups and to look at the clinical phenotypes of this clustering.

METHODS

Study population

Patients who were admitted to the Department of Endocrinology and Metabolism, Shanghai Jiao Tong University Affiliated Sixth People’s Hospital from January 2018 to the end of December 2018 were consecutively enrolled. They were diagnosed with type 2 diabetes according to the 2010 American Diabetes Association standards. This study was a secondary analysis of the CGM and clinical data. For cluster analysis of patients with type 2, a blinded CGM system (iPro2, Medtronic, California, USA) was used in this study. The sensor of the CGM system was inserted on day 0 and removed after 72 hours. The CGM data were calibrated by the fingerstick blood glucose readings no more than every 12 hours. Each participant wore the CGM system for 3 days, and complete data for each participant were extracted on the second day (0:00–24:00). Patients met the following study inclusion criteria: hospitalized patients, age ≥18 years, presence of type 2 diabetes, stable glucose-lowering regimen over the previous 3 months and no data missing in CGM on day 2 and clinical phenotypes data. Exclusion criteria included diabetic ketoacidosis; hyperglycemic hyperosmolar state or severe and recurrent events within the previous 3 months; and history of malignancy or mental disorders. Finally, we succeeded in obtaining complete data for 908 participants. All patients provided written informed consent.

Clinical phenotypes

In our study, each patient underwent physical examination that included measurements of height, weight, and blood pressure. Body mass index (BMI) was calculated as weight (kilograms) divided by squared height (meters). One day before the CGM monitoring period, a venous blood sample was drawn at 06:00 after a 10-hour overnight fast. Fasting C peptide (FCP), HbA1c, high-density lipoprotein cholesterol and low-density lipoprotein cholesterol were determined as previously reported. In addition to FCP, C peptide at 120 min (CP2h) during a standard meal following overnight fast was assayed. The increments of C peptide in plasma levels at 120 min (ΔCP2h=CP2h–FCP) showed the ratio of C peptide response. Homeostasis model assessment (HOMA) indices were added to analyze each cluster according to the HOMA V.2.2.3 calculator.

Multilevel clustering method

The multilevel clustering system was designed as a four-step data-mining cluster model. The first level, features extraction, aimed to extract knowledge-based and statistical-based features from CGM traces. Then Fisher score and variables cluster analyses were applied in the features selection at the second level, aiming at further dimension reduction and obtaining better clustering effect. The three factors from features clustering were put into the third level. The third level, subject clustering, clustered all subjects into different subgroups. The detailed multilevel cluster method follows an iterative procedure, as shown in online supplemental materials section 1.

Features extraction

The 908 CGM traces were first processed to extract the set of well-established indices already considered in the recent research by Weiping et al. As a result, mean sensor glucose (MSG), SD of the sensor glucose (SDSG), coefficient of variation (CV), mean amplitude of glycemic excursions (MAGE), and the largest amplitude of glycemic excursions (LAGE) were calculated into the statistical-based features pool. The knowledge-based features pool had the following features: mean postprandial sensor glucose (MPSG), which means the average sensor value of postprandial glucose 3 hours after three meals; J index and M value; indices based on the permanence in the target glucose range, that is, percentages of values within the target range (TIR; 3.9–10 mmol/L) and percentages of values out of the target range (TOR;
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Table 1 Clinical characteristics of participants with type 2 diabetes

<table>
<thead>
<tr>
<th>Features</th>
<th>Values</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of participants</td>
<td>908</td>
</tr>
<tr>
<td>Gender (male/female)</td>
<td>565/343</td>
</tr>
<tr>
<td>Age (years)</td>
<td>61 (53–67)</td>
</tr>
<tr>
<td>Diabetes duration (years)</td>
<td>12 (6–17)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.89±3.62</td>
</tr>
<tr>
<td>MSG (mmol/L)</td>
<td>8.99 (7.79–10.45)</td>
</tr>
<tr>
<td>MPSG (mmol/L)</td>
<td>10.05 (8.36–11.83)</td>
</tr>
<tr>
<td>TOR (%)</td>
<td>0.34 (0.15–0.55)</td>
</tr>
<tr>
<td>TIR (%)</td>
<td>0.66 (0.45–0.85)</td>
</tr>
<tr>
<td>SDSG (mmol/L)</td>
<td>2.27 (1.60–3.04)</td>
</tr>
<tr>
<td>CV (%)</td>
<td>24.79 (18.80–31.92)</td>
</tr>
<tr>
<td>LAGE (mmol/L)</td>
<td>9.20 (6.70–11.90)</td>
</tr>
<tr>
<td>M value</td>
<td>36.97 (32.41–42.23)</td>
</tr>
<tr>
<td>MAGE (mmol/L)</td>
<td>5.58 (3.99–7.60)</td>
</tr>
<tr>
<td>J index</td>
<td>41.56 (30.06–58.45)</td>
</tr>
<tr>
<td>HBGI</td>
<td>6.38 (3.26–11.10)</td>
</tr>
<tr>
<td>LI</td>
<td>2.50 (1.48–3.79)</td>
</tr>
<tr>
<td>GRADE</td>
<td>373.09±87.06</td>
</tr>
<tr>
<td>ADRR</td>
<td>218.11 (193.01–242.54)</td>
</tr>
</tbody>
</table>

Normally distributed variables are presented as mean±SD, and non-normally distributed data are expressed as median with IQR. ADRR, average daily risk range; BMI, body mass index; CV, coefficient of variation; GRADE, glycemic risk assessment diabetes equation; HBGI, high blood glucose index; LAGE, largest amplitude of glycemic excursions; LI, liability index; MAGE, mean amplitude of glycemic excursions; MPSG, mean postprandial sensor glucose; MSG, mean sensor glucose; SDSG, SD of the sensor glucose; TIR, percentage of values within the target range (3.9–10 mmol/L); TOR, percentage of values out of the target range (<3.9 mmol/L or >10 mmol/L).

Figure 1 Dendrogram of hierarchical cluster analysis of continuous glucose monitoring variables. Height indicates the distance of correlation method between substructures. ADRR, average daily risk range; CV, coefficient of variation; GRADE, glycemic risk assessment diabetes equation score; HBGI, high blood glucose indices; J, J index; LAGE, largest amplitude of glycemic excursions; LI, liability index; M, M value; MAGE, mean amplitude of glycemic excursions; MPSG, mean postprandial sensor glucose; MSG, mean sensor glucose; SDSG, SD of the sensor glucose; TIR, percentages of values within the target range (3.9–10 mmol/L); TOR, percentages of values out of the target range (<3.9 mmol/L or >10 mmol/L).

<3.9 mmol/L or >10 mmol/L); and indices based on glucose risk, that is, low and high blood glucose indices (LBGI, HBGI) and glycemic risk assessment diabetes equation (GRADE) score. In addition, average daily risk range (ADRR) and liability index (LI) were calculated as the feature indices according to the definitions. The ideal blood glucose level in the M value function was set as 5.8 mmol/L.

Through this operation, we reduce the 288-dimensional data of the CGM to a lower dimension of 15. Before the CGM traces were fed into the multilevel clustering, Z-score normalization was used to solve the problem of amplitude scaling and eliminate the offset effect.

Features selection

In the second step, an effective features selection method based on Fisher score and feature clustering were applied. These steps help to find the pattern similarities of CGM features from feature extraction and reducing features for clustering (online supplemental materials section 2). First, the CGM features that have more discrimination information based on Fisher score were selected. Then the agglomerative hierarchical clustering method was adopted for feature fusion. Irrelevant and redundant features were removed. The average of elements in every cluster was calculated and used as a representative value of each cluster in further analysis. Finally, a simpler feature matrix of CGM data was generated.

Subject clustering

In the third step, we applied K-means++ algorithm to the simple features’ matrix from feature selection in order to obtain the subtypes of patients with type 2 diabetes. As an improvement version of K-means, K-means++ is more suitable for large amounts of patient data and its results were more meaningful and easier to interpret. Moreover, the stability and efficiency of the K-means++ method were better than the traditional K-means method by improving the choice of clustering center.

The K-means++ solution with Euclidean distance was designed to identify potential groups among type 2 diabetes based on the simple features’ matrix. We choose the optimal cluster number on the basis of the elbow method. The K-means++ clustering was performed using the cluster function (runs=100) in the scikit-learn package in PYTHON V3.7. Cluster stability was assessed by resampling the data set 2000 times by the Jaccard similarity coefficient.
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Bootstrap method. A stable cluster generally yields Jaccard similarity index of greater than 0.75.\textsuperscript{25}

Statistical analysis
Data were analyzed with PYTHON V.3.7 and PASW Statistics V.25 (SPSS, Chicago, Illinois). To determine the clinical significance of the clustering results, this paper compared the four clusters with the CGM feature matrix and clinical phenotypes. Kruskal-Wallis test was used for non-normally distributed data to determine differences between the four subgroups. A two-tailed \textit{p} value of <0.05 was considered statistically significant.

RESULTS
Participant characteristics
The clinical characteristics are shown in table 1. A total of 908 patients (565 men and 343 women) had a median age of 61 (53–67) years and BMI of 24.89\pm3.62. Two-sample Kolmogorov-Smirnov test indicated that there was no significant difference between sex and all clinical phenotype indices. Therefore, the next step in the subjects’ cluster should not consider sex (online supplemental table S3).

Features cluster
The LBGI was excluded after counting Fisher score. Then the CGM variables were classified into three clusters (figure 1). The first cluster, which was composed of MSG, M value, GRADE, J index, MPSG, HBGI, and TOR, had a high correlation with MSG and HBGI. The second cluster, which was composed of SDSG, CV, LAGE, MAGE, and LI, had the highest correlation with CV. The third cluster was composed of ADRR and TIR. It had a negative correlation with the first and second clusters (online supplemental table S4). Therefore, this study defined the first cluster as the glycemic level factor, the second cluster as the glycemic fluctuation factor and the third cluster as the negative glucose factor.

Subjects cluster
By elbow method, the cluster number of subjects at four would be the most appropriate value (online supplemental materials section 3). Each subgroup has different characteristics on CGM traces throughout follow-up (figure 2). We compared the differences between the clustering factors and selected CGM parameters of the four subgroups (figure 3). Cluster 1, which included 338 (37.22\%) patients, was characterized by the lowest glucose level factor and glucose fluctuation factor and highest negative glucose factor, and was labeled as low-level and low-fluctuation diabetes (LLLFD). Cluster 2, which included 174 (19.16\%) patients and was labeled as high-level and high-fluctuation diabetes (HLHFD), was different from cluster 1; specifically, it had the highest glucose level factor, relatively high glucose fluctuation factor and lowest negative glucose factor. Cluster 3, labeled as the moderate-level and moderate-fluctuation diabetes (MLMFD), included 227 (25.00\%) patients. It...
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was characterized by reasonably low glucose fluctuation factor and high glucose level factor, but otherwise was similar to cluster 1. Overall, 169 (18.61%) patients in cluster 4 (labeled as moderate-level and high-fluctuation diabetes, MLHFD) showed a higher glucose fluctuation factor than those in the other three groups. To be effective clinically, the study compared the clusters regarding clinical phenotypes and antidiabetes agents, as shown in table 2. Cluster 1 (LLLFD) was characterized by the lowest HbA1c (7.2%, 6.5%–8.3%) and highest beta-cell capacity as assessed by C peptide and HOMA-2%β. In contrast, cluster 2 (HLHFD) had the highest HbA1c (10.2%, 9.1%–11.3%) and the most severely impaired beta-cell function. Cluster 3 (MLMFD) had a relatively low HbA1c (8.6%, 7.7%–10.0%) and preserved C peptide indices. Additionally, cluster 3 (MLMFD) and cluster 4 (MLHFD) were comparable (p=0.795) in terms of HbA1c. Cluster 4 showed relatively low levels of beta-cell function indices and a modest level of HbA1c (8.9, 7.5–10.1). However, individuals in cluster 4 (MLHFD) were most sensitive to insulin as reflected by the highest HOMA-2%Sensitivity (3.19, 2.05–53.55) and the lowest HOMA-2%Insulin resistance (3.10, 1.90–4.90). Concerning the treatment received, cluster 1 had the lowest percentage of insulin use (54.4%), while cluster 2 had the highest propensity (93.1%) to receive insulin. Moreover, cluster 4, which is the most insulin-sensitive, had the lowest use of metformin (30.2%) and α-glucosidase inhibitors.

DISCUSSION

This study used a data-driven clustering approach to extract implicit information from extensive high-dimensional CGM data. Distinct from previous efforts, this is a method that could not only provide us with those knowledge-based and statistics-based CGM indices, but also offer features selection to improve the clustering results of subjects. It may provide new insights into the clustering of type 2 diabetes and more tailored diabetes management. The results demonstrate that this multi-level clustering approach could identify patients with type 2 diabetes with diverse clinical phenotypes, such as beta-cell function. Some medical check-ups like C peptide testing is not routinely measured, and glucose pattern represents the most obvious phenotypes of diabetes and may reveal the inherent nature of the disease. Furthermore, the method can be applied to CGM sensors for the convenience of patients with type 2 diabetes at home. The robustness of the clustering was confirmed by the distinct characteristics of clinical phenotypes, particularly for insulin secretion/sensitivity indices, among the four subgroups. In short, cluster 1 (LLLFD) and cluster 2 (HLHFD) were related to the best and the worst beta-cell function, respectively. Intriguingly, while beta-cell function was modestly impaired, patients in cluster 4 (MLHFD), which was characterized by unstable glucose levels, had the highest level of insulin sensitivity, highlighting the role of insulin sensitivity in glucose homeostasis. Consistent with our study, a positive relationship between insulin sensitivity and glycemic variability was reported in 366 insulin-treated hospitalized patients with type 2 diabetes. Indeed, from the clinical perspective, those who are more insulin-sensitive may be more prone to glycemic excursions after dose adjustment. Given the distinct clinical features among the four clusters, different strategies of glucose-lowering treatment may be considered. For instance, for cluster 2, with the most impaired beta-cell function, insulin therapy should be initiated at the early stage of the disease to achieve glycemic targets. For patients in cluster 4, antidiabetic agents that alleviate glucose fluctuations, such as glucagon-like peptide 1 receptor agonists, should be preferentially considered.

Figure 3  Box plot of cluster factors in patients. Adjusted p value was used. **Adjusted p values were under 0.01; ***adjusted p values were under 0.001. HLHFD, high-level and high-fluctuation diabetes; LLLFD, low-level and low-fluctuation diabetes; MLHFD, moderate-level and high-fluctuation diabetes; MLMFD, moderate-level and moderate-fluctuation diabetes.
## Table 2 Clinical characteristics of the various clusters of type 2 diabetes

<table>
<thead>
<tr>
<th>Clinical phenotypes</th>
<th>Clusters</th>
<th>LLLFD (n=338)</th>
<th>HLHFD (n=174)</th>
<th>MLMFD (n=227)</th>
<th>MLHFD (n=169)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Basic information</strong></td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td></td>
<td>62 (54–68)</td>
<td>61 (51–67)</td>
<td>60 (53–67)</td>
<td>62 (54–67)</td>
<td>0.166</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td></td>
<td>24.23 (22.49–26.63)</td>
<td>24.94 (22.65–27.26)</td>
<td>24.80 (22.82–27.31)</td>
<td>24.22 (22.56–26.36)</td>
<td>0.150</td>
</tr>
<tr>
<td>Duration (years)</td>
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<td>12 (6–17)</td>
<td>14 (7–20)</td>
<td>11 (6–18)</td>
<td>12 (5–17)</td>
<td>0.154</td>
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<tr>
<td><strong>HOMA indices</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HOMA-2%β</td>
<td></td>
<td>172.45 (118.08–239.40)</td>
<td>73.20* (50.95–106.95)</td>
<td>115.70*† (82.70–155.50)</td>
<td>110.90*† (71.75–172.00)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>HOMA-2% Sensitivity</td>
<td></td>
<td>23.40 (16.60–32.83)</td>
<td>26.25 (16.58–38.93)</td>
<td>21.80 (14.90–33.60)</td>
<td>31.90*†‡ (20.50–53.55)</td>
<td>&lt;0.001</td>
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<tr>
<td>HOMA-2% Insulin resistance</td>
<td></td>
<td>4.30 (3.00–6.00)</td>
<td>3.80 (2.58–6.03)</td>
<td>4.60 (3.00–6.70)</td>
<td>3.10*†‡ (1.90–4.90)</td>
<td>&lt;0.001</td>
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<tr>
<td><strong>Clinical measures</strong></td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td></td>
<td>7.2 (6.4–8.3)</td>
<td>10.2* (9.1–11.3)</td>
<td>8.6† (7.7–10.0)</td>
<td>8.9† (7.5–10.1)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>FCP (ng/mL)</td>
<td></td>
<td>1.84 (1.30–2.53)</td>
<td>1.43* (0.97–2.14)</td>
<td>1.73† (1.21–2.62)</td>
<td>1.26*‡ (0.76–1.98)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>CP2h (ng/mL)</td>
<td></td>
<td>4.47 (2.73–6.56)</td>
<td>2.43* (1.73–3.88)</td>
<td>4.31† (2.35–6.17)</td>
<td>3.29*† (1.89–5.38)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>ΔCP2h (ng/mL)</td>
<td></td>
<td>2.65 (1.08–4.38)</td>
<td>1.35* (0.55–2.17)</td>
<td>2.21† (0.88–3.83)</td>
<td>1.89† (0.93–3.41)</td>
<td>&lt;0.001</td>
</tr>
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<td>SBP (mm Hg)</td>
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<td>130 (120–140)</td>
<td>130 (120–140)</td>
<td>130 (120–140)</td>
<td>130 (120–140)</td>
<td>0.620</td>
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<tr>
<td>DBP (mm Hg)</td>
<td></td>
<td>78 (70–82)</td>
<td>80 (70–84)</td>
<td>80 (70–84)</td>
<td>78 (70–82)</td>
<td>0.793</td>
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<tr>
<td>Triglyceride (mmol/L)</td>
<td></td>
<td>1.32 (0.92–1.88)</td>
<td>1.55* (1.12–2.80)</td>
<td>1.67* (1.13–2.50)</td>
<td>1.27† (0.97–1.94)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td></td>
<td>1.00 (0.84–1.23)</td>
<td>0.96 (0.82–1.15)</td>
<td>0.97 (0.80–1.17)</td>
<td>1.06†† (0.90–1.25)</td>
<td>0.008</td>
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<tr>
<td>LDL cholesterol (mmol/L)</td>
<td></td>
<td>2.51 (1.96–3.22)</td>
<td>2.65 (2.06–3.23)</td>
<td>2.55 (1.95–3.30)</td>
<td>2.75 (1.98–3.35)</td>
<td>0.286</td>
</tr>
<tr>
<td><strong>Antidiabetic agents, n (%)</strong></td>
<td></td>
<td>Metformin 153 (45.3)</td>
<td>61 (35.1)</td>
<td>97 (42.7)</td>
<td>51† (30.2)</td>
<td>0.004</td>
</tr>
<tr>
<td>Sulfonylurea 83 (24.6)</td>
<td>22* (12.6)</td>
<td>63† (27.8)</td>
<td>25† (13.0)</td>
<td>&lt;0.001</td>
<td></td>
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<tr>
<td>Thiazolidinediones 18 (5.3)</td>
<td>5 (2.9)</td>
<td>15 (6.6)</td>
<td>8 (4.7)</td>
<td>0.400</td>
<td></td>
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<tr>
<td>Glinides 20 (5.9)</td>
<td>5 (2.9)</td>
<td>14 (6.2)</td>
<td>11 (6.5)</td>
<td>0.400</td>
<td></td>
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<tr>
<td>DPP-4 inhibitors 32 (9.5)</td>
<td>9 (5.2)</td>
<td>30† (13.2)</td>
<td>11 (6.5)</td>
<td>0.025</td>
<td></td>
<td></td>
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<tr>
<td>AGI 132 (39.1)</td>
<td>48* (27.6)</td>
<td>86 (37.9)</td>
<td>29† (17.2)</td>
<td>&lt;0.001</td>
<td></td>
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</tr>
<tr>
<td>Insulin 184 (54.4)</td>
<td>162* (93.1)</td>
<td>157† (69.2)</td>
<td>143†† (84.6)</td>
<td>&lt;0.001</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

All data are expressed as median with IQR.

For clusters, p values are from Kruskal-Wallis test for continuous variables and from χ² for categorical variables.

Duration refers to duration of diabetes.

*Significant difference in unpaired (p<0.05), Dunn-Bonferroni test for post-hoc comparisons, compared with cluster 1.
†Significant difference in unpaired (p<0.05), Dunn-Bonferroni test for post-hoc comparisons, compared with cluster 2.
‡Significant difference in unpaired (p<0.05), Dunn-Bonferroni test for post-hoc comparisons, compared with cluster 3.

AGI, α-glucosidase inhibitor; BMI, body mass index; CP2h, C peptide levels at 2 hours; ΔCP2h, increments of C peptide in plasma levels 120 min; DBP, diastolic blood pressure; DPP-4, Dipeptidyl peptidase 4; FCP, C peptide variables as fasting C peptide; HbA1c, glycated hemoglobin A1c; HDL, high-density lipoprotein; HLHFD, high-level and high-fluctuation diabetes; HOMA, Homeostasis model assessment; LDL, low-density lipoprotein; LLLFD, low-level and low-fluctuation diabetes; MLHFD, moderate-level and low-fluctuation diabetes; MLMFD, moderate-level and moderate-fluctuation diabetes; SBP, systolic blood pressure.
Another aspect of clinical significance is the presumed differential risk for chronic diabetic complications across the four clusters. Cluster 2 (HLHFD) and cluster 4 (MLHFD) are more associated with hyperglycemia because their glucose fluctuated wildly. In addition, the high degree of glycemic fluctuation factor noted in these two clusters may confer additional risk for microvascular and macrovascular complications. Cluster 3 (MLMFD), with relatively higher insulin resistance, may have a high risk of kidney disease due to the correlation between insulin resistance and glomerular hypertension and hyperfiltration. It may help doctors to choose precise treatment more quickly according to cluster phenotypes.

The limitations of this study are worth noting. First, the abnormal samples in the database were not removed by our approach. This may reduce the stability of clustering results. In future studies, the method should be improved to exclude outlier subjects. Second, our study only included hospitalized patients with type 2 diabetes in China. The generalizability of the study results is uncertain and needs to be tested in other populations. Third, it is notable that some factors could affect the stability of glucose profile over time within individuals, such as diet, exercise, stress and treatment regimen. In addition, only 1-day CGM data were used in our study, while two previous studies suggested that 14 days of CGM may be needed for reliable estimation of overall glucose control. Therefore, the results of CGM clustering should be interpreted with caution. Finally, whether the new method of clustering translates into improved glycemic control and subsequent diabetes-related outcomes remains unknown. Further studies are warranted to validate the clinical significance of this method.

In summary, a multilevel clustering method driven by CGM data was designed to divide patients with type 2 diabetes into subgroups with distinct clinical phenotypes. This method has implications for big data from CGM systems and for digital precision in diabetes research. Furthermore, the results of the study show the promise of developing personalized medicine and intelligent healthcare for type 2 diabetes.

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Contributors RT, XY and JZ designed the study. RT was responsible for data analysis and overall design of the algorithm. XY was responsible for design and analysis of the algorithm. JZ and JL were responsible for clinical phenotypes analysis. JL, YS, WL, WZ and YB assisted in data collection and offered suggestion for the study. HL and JZ funded the study. All authors read and approved the final manuscript.

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Competing interests None declared.

Patient consent for publication Not required.

Ethics approval The study was approved by the Ethics Committee of Shanghai Jiao Tong University Affiliated Sixth People’s Hospital (2020-KY-024) and complied with the principles of the Helsinki Declaration.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement Data are available upon reasonable request.

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A Multilevel Clustering Approach Driven by Continuous Glucose Monitoring Data for Further Classification of Type 2 Diabetes

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1. **The overview of multilevel clustering of CGM data:**

The multilevel clustering method as a data-mining model contains four steps. The first step extracts features from CGM data. Statistic-base features like mean values (mean sensor glucose, MSG), standard deviation of the values (SD of the sensor glucose, SDBG) and coefficient of variation (CV) were calculated. In addition, some knowledge-base features which are used in clinical were also extracted. Globally, a 15-dimensional group of feature variables extracted from time-series CGM data was used to characterize the patient for future analysis. Then we designed a step for selecting features in the feature group. In this step, an effective feature selection method based on Fisher score and agglomerative hierarchical clustering was applied to find the pattern similarities of CGM features and reducing features. Through this step, the original 15-dimensional features could be further reduced to 3-dimensional features with clinical significance. In the third step, we used the 3-dimensional features in the second step to characterize the participants. And subgroups of the participants were found by K-means++ method. At the last step, four subgroups were identified and their clinical phenotypes were compared. The overall flow chart is shown in [Supplementary Figure S1](#). Furthermore, the details of the method are shown in [Supplementary Table S1](#).
Supplementary Figures S1: The flow chart of the cluster method.

Abbreviations: LBGI-low blood glucose indices, K-W test-Kruskal Wallis test, AHC-agglomerative hierarchical clustering
Supplementary Table S1: The iterative procedure of Multilevel cluster for CGM data

Algorithm Multilevel cluster for CGM data (MLC-CGM)

**Input:** CGM datasets

**Output:** the groups’ labels of each subject

**Step 1: Feature Extraction**
   **Step 1.1:** Calculate knowledge-based features
   **Step 1.2:** Calculate statistical-base features
   **Step 1.3:** Initialize the set of features $S$

**Step 2: Feature Selection**
   **Step 2.1:** Build a simple clustering using $S$ as input
   **Step 2.2:** For every feature $f_i$ in $S$:
      **Step 2.2.1:** Compute the Fisher Score ($FS$).
      **Step 2.2.2:** Find $f^* = f_i$ if $FS > \text{threshold}$ such that adding it to $S$ set=$S$
   **Step 2.3:** Update the $S^*$ set
   **Step 2.4:** Cluster features in $S^*$ set

**Step 3: Subjects Cluster**
   **Step 3.1:** Choose the optimal clustering number $n$ by elbow method
   **Step 3.2:** Cluster patients into $n$ subgroups among $S^*$ set

**Step 4:** Definite the subgroups using Kruskal-Wallis test
2. Fisher score

As a state-of-the-art feature selection, the main idea of the Fisher score is that the features with strong discrimination performance keep the distance within clusters as small as possible and keep the distance between clusters as large as possible. Compared to Mutual information (MI), Relief, Laplacian Score and Trace ratio criterion, Fisher Score (FS) is one of the most widely used criteria for filter-based feature selection. Here, the Fisher score of a single feature, used as the criterion for selecting the CGM indices can be expressed by the following formulas:

\[
J_{\text{fisher}}(k) = \frac{S_B(k)}{S_w(k)}
\]

\[
S_B(k) = \sum_{i=1}^{c} \frac{n_i}{n} \left( m_i(k) - m(k) \right)^2
\]

\[
S_w(k) = \sum_{i=1}^{c} \sum_{x \in S_i} \left( x(k) - m_i(k) \right)^2
\]

where there are \( n \) samples in the data set that belong to \( C \) clusters, each cluster contains \( n_i \) samples respectively. \( x(k), m_i(k), m(k) \) means the sample \( x \), mean values of the \( i \) cluster, and all samples' mean values on the \( k \) features. The Fisher score under the threshold will be removed. Here, we set the threshold value to 0.5. And the Fisher score of the CGM features are shown in Supplementary Tables S2.

### Supplementary Table S2. The fisher score results in the feature selection

<table>
<thead>
<tr>
<th>Feature</th>
<th>FS values</th>
<th>Features</th>
<th>FS values</th>
<th>Features</th>
<th>FS values</th>
</tr>
</thead>
<tbody>
<tr>
<td>MSG</td>
<td>3.23</td>
<td>CV</td>
<td>0.92</td>
<td>LBGI</td>
<td>0.22</td>
</tr>
<tr>
<td>MPSG</td>
<td>2.53</td>
<td>LAGE</td>
<td>1.44</td>
<td>HBGI</td>
<td>3.75</td>
</tr>
<tr>
<td>TIR</td>
<td>3.44</td>
<td>M value</td>
<td>2.98</td>
<td>ADRR</td>
<td>2.08</td>
</tr>
<tr>
<td>TOR</td>
<td>3.44</td>
<td>MAGE</td>
<td>1.00</td>
<td>GRADE</td>
<td>2.37</td>
</tr>
<tr>
<td>SDSG</td>
<td>1.46</td>
<td>J-index</td>
<td>3.56</td>
<td>LI</td>
<td>0.57</td>
</tr>
</tbody>
</table>

**Abbreviations:** MSG-mean sensor glucose; MPSG-mean postprandial sensor glucose; TIR-percentages of values within the target range ([3.9-10] mmol/L); TOR-percentages of values out of the target range (<3.9 or >10 mmol/L); SDSG-standard deviation of the sensor glucose, CV-coefficient of variation; LAGE-largest amplitude of glycemic excursions; MAGE-mean amplitude of glycemic excursions; HBGI,LBGI-high or low blood glucose indices; GRADE-glycemic risk assessment diabetes equation score; LI-liability index; ADRR-average daily risk range.
3. Elbow method

To properly choose the optimal number of the K-means++ on the basis of the elbow method, we divided the data into categories 1 to 11. Then the error sum of the square distance (SSE) between the particle of each cluster and the sample point in the cluster were calculated as:

$$SSE = \sum_{i=1}^{k} \sum_{x \in C_i} \|x - u_i\|^2$$

(2)

where $u_i$ is the center of cluster $i$, $x$ is the sample point, and $k$ is the value of K-means++. Distortions of the patient’s matrix at 4 will be the most appropriate values as shown in Supplementary Figures S2.

Supplementary Figure S2. The SSE line chart of the elbow method. The curve reached the break point at $k=4$. As $k$ increases, SSE decreases slowly.
Supplementary Table S3. Two sample Kolmogorov-Smirnov test results

<table>
<thead>
<tr>
<th>SEX</th>
<th>AGE</th>
<th>BMI</th>
<th>Duration</th>
<th>SBP</th>
<th>DBP</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.000</td>
<td>0.001</td>
<td>0.109</td>
<td>0.067</td>
<td>0.158</td>
</tr>
<tr>
<td>SEX</td>
<td>HbA1c</td>
<td>HDL</td>
<td>LDL</td>
<td>FCP</td>
<td>CP2h</td>
</tr>
<tr>
<td></td>
<td>0.003</td>
<td>0.000</td>
<td>0.025</td>
<td>0.145</td>
<td>0.175</td>
</tr>
</tbody>
</table>

A two-tailed P value of <0.010 was considered statistically significant. As shown in table, most adjusted P values above 0.010. These results verified the similar distributions at characteristics in men and women. So, sex could not be considered in the analysis.

Abbreviations: FCP-C peptide variables as fasting C peptide; CP2h-C peptide levels at 2 hours; ΔCP120min- the increments of C peptide in plasma levels 90 minutes; SBP-systolic blood pressure; DBP-diastolic blood pressure; BMI-body mass index; Duration-duration of diabetes; HDL-high density lipoprotein; LDL-low density lipoprotein.

Supplementary Table S4. Glycemic level factor (GLF), the glycemic fluctuation factor (GFF) and negative glycemic factor (NGF) correlation analysis

<table>
<thead>
<tr>
<th></th>
<th>MSG</th>
<th>HBGI</th>
<th>CV</th>
<th>TIR</th>
</tr>
</thead>
<tbody>
<tr>
<td>GLF</td>
<td>0.988</td>
<td>0.993</td>
<td>0.120</td>
<td>-0.925</td>
</tr>
<tr>
<td>GFF</td>
<td>0.365</td>
<td>0.530</td>
<td>0.899</td>
<td>-0.553</td>
</tr>
<tr>
<td>NGF</td>
<td>-0.986</td>
<td>-0.980</td>
<td>-0.047</td>
<td>0.935</td>
</tr>
</tbody>
</table>

A two-tailed P value of <0.010 was considered statistically significant. And the number with a * means its P value above 0.010. As shown in the table (Table S2), GLF had a higher correlation with MSG and HBGI, and GFF had the highest correlation with CV. NGF had the negative correlation with the indices unlike Factor 1 and Factor 2.

Abbreviations: MSG-mean sensor glucose; HBGI- high blood glucose indices; CV-coefficient variation; TIR-time in range [3.9-10] mmol/L.
References: