


Insulin resistance and liver histopathology in metabolically unhealthy subjects do not correlate with the hepatic abundance of NLRP3 inflammasome nor circulating IL-1 β levels

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To cite: Quezada N, Valencia I, Torres J, *et al.* Insulin resistance and liver histopathology in metabolically unhealthy subjects do not correlate with the hepatic abundance of NLRP3 inflammasome nor circulating IL-1 β levels. *BMJ Open Diab Res Care* 2021;**9**:e001975. doi:10.1136/bmjdr-2020-001975

► Supplemental material is published online only. To view, please visit the journal online (<http://dx.doi.org/10.1136/bmjdr-2020-001975>).

Received 31 October 2020
Revised 17 March 2021
Accepted 3 April 2021



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ABSTRACT

Introduction Systemic chronic low-grade inflammation has been linked to insulin resistance (IR) and non-alcoholic steatohepatitis (NASH). NOD-like receptor protein 3 (NLRP3) inflammasome and its final product, interleukin (IL)-1 β , exert detrimental effects on insulin sensitivity and promote liver inflammation in murine models. Evidence linking hepatic NLRP3 inflammasome, systemic IR and NASH has been scarcely explored in humans. Herein, we correlated the hepatic abundance of NLRP3 inflammasome components and IR and NASH in humans.

Research design and methods Metabolically healthy (MH) (n=11) and metabolically unhealthy (MUH) (metabolic syndrome, n=21, and type 2 diabetes, n=14) subjects were recruited. Insulin sensitivity (homeostatic model assessment of IR (HOMA-IR) and Oral Glucose Sensitivity (OGIS₁₂₀)), glycemic (glycated hemoglobin), and lipid parameters were determined by standard methods. Plasma cytokines were quantified by Magpix. Hepatic NLRP3 inflammasome components were determined at the mRNA and protein levels by reverse transcription-quantitative PCR and western blot, respectively. Liver damage was assessed by histological analysis (Non-alcoholic Fatty Liver Disease Activity Score (NAS) and Steatosis, Inflammatory Activity, and Fibrosis (SAF) scores). IR and liver histopathology were correlated with NLRP3 inflammasome components as well as with liver and plasma IL-1 β levels.

Results Body Mass Index, waist circumference, and arterial hypertension frequency were significantly higher in MUH subjects. These patients also had increased high-sensitivity C reactive protein levels compared with MH subjects. No differences in the plasma levels of IL-1 β nor the hepatic content of *Nlrp3*, apoptosis-associated speck-like (*Asc*), *Caspase-1*, and *IL-1 β* were detected between MUH and MH individuals. MUH subjects had significantly higher NAS and SAF scores, indicating more severe liver damage. However, histological severity did not correlate with the hepatic content of NLRP3 inflammasome components nor *IL-1 β* levels.

Conclusion Our results suggest that NLRP3 inflammasome activation is linked neither to IR nor to the inflammatory status of the liver in MUH patients.

Significance of this study

What is already known about this subject?

- NLRP3 inflammasome expression in adipose tissue has been linked to insulin resistance (IR) in murine models and humans.
- Human liver NLRP3 inflammasome expression has been studied in primary liver diseases.
- No studies have ever linked hepatic NLRP3 inflammasome expression with IR and non-alcoholic fatty liver disease severity in humans.

What are the new findings?

- Hepatic NLRP3 inflammasome expression is linked neither to human IR determined by the National Cholesterol Education Program Adult Treatment Panel III criteria nor to type 2 diabetes mellitus.
- Hepatic NLRP3 inflammasome expression is not associated with the severity of liver pathology in non-alcoholic fatty liver disease.

How might these results change the focus of research or clinical practice?

- Quantification of hepatic NLRP3 inflammasome should not be the focus for assessing IR nor non-alcoholic fatty liver disease in humans.

INTRODUCTION

Chronic low-grade systemic inflammation has been involved in the pathogenesis of insulin resistance (IR).^{1–3} In fact, dysregulation of innate immunity is a possible cause of hepatic IR as well as hepatitis in non-alcoholic fatty liver disease (NAFLD).^{4–9}

Innate immunity is triggered by a variety of pathogen and host-derived molecules, including bacterial lipopolysaccharide, single-stranded or double-stranded RNA, cholesterol and uric acid crystals, among others. These ligands activate molecular pattern

recognition receptors at the cell surface and cytosol. Inflammasomes are multiprotein cytosolic molecular pattern complexes that catalyze the proteolytic conversion of prointerleukin (IL)-1 β into mature IL-1 β .^{10–14} IL-1 β is a potent proinflammatory cytokine that decreases insulin-stimulated glucose uptake in cultured adipocytes.^{15–17} Notably, pharmacological blocking of the IL-1 receptor with anakinra improves glycemic control in patients with type 2 diabetes (T2D),^{18,19} suggesting a direct link between IL-1 β signaling and IR. Finally, IL-1 β may also drive islet β -cell dysfunction and thus promote T2D.^{20,21}

The role of inflammasome on IR has been mostly substantiated by studies in animal models of obesity, diabetes, and NAFLD.^{22–25} In rodents, nod-like receptor protein 3 (NLRP3) inflammasome gene deletion generates resistance to diet-induced obesity and improves IR in mice.²⁶ Additionally, transgenic overexpression of specific components of NLRP3 inflammasome increases liver inflammation and fibrosis, and, conversely, *Nlrp3* gene deletion ameliorates hepatic steatosis and inflammation in models of diet-induced NAFLD.^{27–29}

The level of evidence in human studies is much lower. The role of *Nlrp3* in obesity, IR, and T2D has been evaluated mostly at the adipose tissue level, suggesting a positive correlation between *Nlrp3* gene expression and IR.^{25,30} Additionally, hepatic levels of *Nlrp3* and IL-1 β mRNA are reportedly increased in patients with non-alcoholic steatohepatitis (NASH) compared with patients with pure steatosis, suggesting a role for NLRP3 inflammasome in NAFLD progression.²⁹

Nevertheless, the pathophysiological relevance of hepatic NLRP3 inflammasome on insulin sensitivity as well as in liver inflammation has been assessed only in extremely obese subjects, remaining uncertain whether NLRP3 inflammasome abundance and activity correlate with IR in less obese individuals.²⁹

In this work, we aimed to assess whether the hepatic abundance of NLRP3 inflammasome components correlates with systemic markers of inflammation, IR, and liver histological alterations in a cohort of metabolically healthy (MH) and metabolically unhealthy (MUH) subjects.

RESEARCH DESIGN AND METHODS

Selection of patients and ethical considerations

This study accomplished the Strengthening the Reporting of Observational Studies in Epidemiology guidelines (<https://www.strobe-statement.org/>). All subjects were evaluated in the digestive surgery outpatient clinic of Pontificia Universidad Católica de Chile School of Medicine between July 2015 and December 2018. Their original consultations were benign abdominal diseases requiring open or laparoscopic surgery, including gastroesophageal reflux disease, cholelithiasis, obesity, and abdominal wall hernias. Subjects were invited to participate after evaluation of the status of systemic IR according

to National Cholesterol Education Program Adult Treatment Panel III (ATP III) criteria for metabolic syndrome (MS), with waist circumference cut-off values adjusted to the Chilean population.^{31–33} Subjects of both sexes, aged between 18 and 60 years old, were included. This cohort was separated into two groups: (1) MH subjects, corresponding to those with 0 or 1 criteria for MS, but without T2D; and (2) MUH subjects, corresponding to non-diabetic patients with three to five ATP III criteria, and patients with T2D,³⁴ irrespective of their MS status. Individuals with two criteria for MS were excluded because they have an intermediate metabolic status of uncertain pathophysiological meaning.

Patients were invited to participate if they met the described inclusion criteria and the following exclusion criteria: (1) current or past oncological diseases; (2) chronic inflammatory systemic diseases (systemic erythematous lupus, rheumatoid arthritis, and inflammatory bowel disease); (3) chronic infections (HIV, human hepatitis virus B and C, tuberculosis, etc); (4) pregnancy; (5) chronic organ failure syndromes (chronic obstructive lung disease, chronic liver and renal disease, and congestive cardiac failure); (6) patients with T2D under insulin therapy; and (7) subjects with clinically significant alcohol consumption (≥ 20 g/day for women and ≥ 30 g/day for men). Exclusion criteria were aimed to prevent conditions that may activate chronic systemic inflammatory mechanisms, induce a different pathway of liver damage, or trigger insulin secretion deficiency or IR by independent mechanisms.

All patients underwent a preoperative evaluation consisting of physical examination (body weight, height, waist circumference, and arterial blood pressure determination) and venous blood draw to determine biochemical markers of MS and systemic inflammation. Liver samples were harvested on the day of the elective surgeries, and no follow-up was performed after surgery for this study.

Laboratory analysis

Complete blood cell count, liver enzymes, lipid profile, glucose, and insulin levels, A1c glycated hemoglobin, thyroid function tests, and highly sensitive C reactive protein (hsCRP) were determined in the central laboratory of Christus Health–Pontificia Universidad Católica de Chile network. Dynamic insulin sensitivity was assessed by an oral glucose tolerance test (75 g) with multiple venous blood sampling to determine glucose and insulin levels and calculate the Oral Glucose Sensitivity (OGIS₁₂₀) index,³⁵ a validated surrogate for clamping procedures. All tests were performed after 10–12 hours of overnight fasting, between 09:00 and 10:00 in the morning. Plasma and serum samples were stored at -80°C for subsequent analyses.

Plasma levels of IL-1 β , tumor necrosis factor alpha (tnf- α), IL-6, macrophage chemoattractant protein-1, resistin, plasminogen activator inhibitor-1 (pai-1), adiponectin, hepatocyte growth factor, IL-8, nerve growth factor, and

Table 1 Anthropometrics, clinical status, and laboratory tests of the study population

Group (n)	Healthy controls	Metabolic syndrome	T2D	P value
	ATP III 0–1 (n=11)	ATP III 3–5 (n=21)	Any ATP III (n=14)	
Female gender, n (%)	9 (82)	14 (67)	7 (50)	0.24*
Age (years) (mean±SD)	41±10	43±9,6	48±8,8	0.17
Body mass index (kg/m ²) (mean±SD)	29.7±8	38.2±7.5	37.6±5.3	0.0066*†‡
Waist circumference (cm) (mean±SD)	87±14	105±13	106±13	0.0009*†‡
Hypertension, n (%)	0 (0)	12 (50)	5 (36)	0.01*
Hypercholesterolemia, n (%)	2 (18)	12 (57)	9 (64)	0.05*
Statins (%)	0/2 (0)	1/12 (8)	7/9 (78)	0.006*
Glycemia (mg/dL) (mean±SD)	83±6	92±8	118±32	<0.0001‡§
Insulinemia (U/mL) (mean±SD)	9.7±3.6	17.9±7.8	20±10.5	0.0069†‡
HOMA-IR (mean±SD)	1.99±0.8	4±1.7	5.8±3.2	0.003†‡§
HbA1c (%) (mean±SD)	5.1±0.3	5.6±0.3	6.7±1.2	<0.0001‡§
OGIS ₁₂₀ (mean±SD)	444±56	330±71	312±52	<0.0001†‡
Total cholesterol (mg/dL) (mean±SD)	177±32	197±54	168±43	0.18
HDL cholesterol (mg/dL) (mean±SD)	54±13	41±11	42±11	0.006†‡
LDL cholesterol (mg/dL) (mean±SD)	105±30	123±44	96±36	0.12
Triglycerides (mg/dL) (mean±SD)	86±26	166±82	175±102	0.01†‡
ATP III criteria (mean±SD)	0.7±0.47	3.6±0.59	3.6±0.76	<0.001†‡
hsCRP (mg/L) (mean±SD)	1.3±0.6	6.6±4.3	6.7±4.7	0.01†‡

Statistical analysis performed with one-way analysis of variance and Tukey post-test to compare pairs of groups.

*Denotes χ^2 statistical analysis. Significant difference was considered when the p value is <0.05.

†Denotes significant difference between ATP III 0–1 vs ATP III 3–5.

‡Denotes significant difference between ATP III 0–1 vs T2D.

§Denotes significant difference between ATP III 3–5 vs T2D.

ATP III, Adult Treatment Panel III; HbA1c, glycated hemoglobin; HDL, high-density lipoprotein; HOMA-IR, homeostatic model assessment of insulin resistance; hsCRP, high-sensitivity C reactive protein; LDL, low-density lipoprotein; OGIS₁₂₀, Oral Glucose Sensitivity; T2D, type 2 diabetes.

leptin were measured with MAGPIX system using the HADCYMAG-61K kit (Luminex®).

Liver tissue sampling

On laparoscopy, a wedge liver tissue sample was taken from the lateral liver segment with cold scissors. Tru-cut needle-based biopsies are no longer allowed in the Department of Digestive Surgery of Pontificia Universidad Católica de Chile because of reported severe intraoperative and post-operative hemorrhagic complications.³⁶ Furthermore, excisional liver biopsies have comparable morphological and molecular qualities to deep puncture biopsies when examined by an expert pathologist aware of the excessive fibrosis that occurs near Glisson's capsule. To further control the histological quality of the biopsies included in this study, only samples with at least 11 complete portal tracts were analyzed.^{37–39} Liver samples were immediately separated into three pieces: one was stored in RNAlater storage solution; a second was snap-frozen in liquid nitrogen and stored at –80°C; and a third was fixed in formaldehyde for histological analyses.

Western blot analysis

Liver tissue was homogenized in ice-cold RIPA buffer (50 mM, Tris-Cl, pH 7.5, 150 mM NaCl, 1% NP-40, 0.5% sodium deoxycholate, and 1% sodium dodecyl sulfate), supplemented with a protease and phosphatase inhibitor cocktail tablet (Roche Diagnostics, GmbH). Protein concentration was determined with BCA Protein Assay Kit (Pierce Biotechnology, Rockford, Illinois, USA). Forty micrograms of proteins were resolved in 10% sodium dodecyl sulfate–polyacrylamide gel electrophoresis and transferred onto polyvinylidene fluoride or nitrocellulose membranes. Primary antibodies against NLRP3 (monoclonal antibody, Cell Signaling Technology), ASC (Cell Signaling Technology), and caspase-1 (Cell Signaling Technology) were used. A horseradish peroxidase-linked secondary antibody anti-rabbit IgG (Cell Signaling Technology) was used. β -Actin or β -tubulin was used as loading control (Cell Signaling Technology).

Reverse transcription–quantitative PCR (RT-qPCR)

Total RNA was extracted from 100 mg of frozen liver tissue, using the PureLink RNA Mini Kit (Invitrogen), treated with

Table 2 Plasma cytokine levels of healthy controls and subjects with metabolic syndrome and T2D

Group (n)	Healthy controls	Metabolic syndrome	T2D	P value
	ATP III 0–1 (n=13)	ATP III 3–5 (n=24)	Any ATP III (n=13)	
Adiponectin (pg/mL) (mean±SD)	51.9±35.6	15.6±9.2	20±14.9	0.001*†
Resistin (pg/mL) (mean±SD)	35.3±14.5	44.6±15.7	36.6±11.8	0.15
PAI-1 (pg/mL) (mean±SD)	32.9±14.7	56±21.9	55.23±17.1	0.005*†
NGF (pg/mL) (mean±SD)	1.8±0.6	2.2±0.8	3.1±2.1	0.058
IL-6 (pg/mL) (mean±SD)	3.1±1.3	6.9±4.7	7.1±5.3	0.03*
Leptin (pg/mL) (mean±SD)	31.7±26.3	52.6±33.3	45.3±25.5	0.17
IL-8 (pg/mL) (mean±SD)	6.5±3.5	5.6±2.1	6.1±3.1	0.66
HGF (pg/mL) (mean±SD)	185.5±95.9	263.3±117.2	321.3±119.2	0.01†
MCP-1 (pg/mL) (mean±SD)	162±30.8	219.2±84	204.2±73.8	0.13
TNF- α (pg/mL) (mean±SD)	2.4±0.75	3.52±1.52	3.46±0.83	0.03*
IL-1 β (pg/mL) (mean±SD)	0.68±0.25	0.79±0.35	0.8±0.37	0.62

Statistical analysis performed with one-way analysis of variance and Tukey post-test to compare pairs of groups. Significant difference was considered when the p value was <0.05.

*Denotes significant difference between ATP III 0–1 vs ATP III 3–5

†Denotes significant difference between ATP III 0–1 vs T2D.

ATP III, Adult Treatment Panel III; HGF, hepatocyte growth factor; IL, interleukin; MCP-1, macrophage chemoattractant protein-1; NGF, nerve growth factor; PAI-1, plasminogen activator inhibitor-1; T2D, type 2 diabetes; TNF- α , tumor necrosis factor alpha.

DNase, and reverse transcribed. RT-qPCR was performed according to the protocol provided by TaqMan Universal PCR Master Mix (Applied Biosystems). Amplification curves and specific product detection were quantified and analyzed with ABI PRISM Step One Fast Sequence Detection System (Applied Biosystems). Primers used for reactions were *Nlrp3* (Hs00918082_m1), *Caspase-1* (Hs00354836_m1), and *IL-1 β* (Hs00174097_m1). Relative mRNA abundances were determined with the delta–delta cycle threshold (Ct) method using β -actin (Hs01060665_g1) as a housekeeping gene, as previously described^{40–42}

Liver histology

Liver tissue samples were fixed in 10% buffered formalin, embedded in paraffin, and sectioned with a microtome. Tissue sections were stained with H&E and Van Gieson staining. Surgical pathologists, blind for the clinical status of the patients, evaluated the specimens and assigned a score for steatosis, inflammation, ballooning, and fibrosis. Steatosis, Inflammatory Activity, and Fibrosis (SAF) score and Non-alcoholic Fatty Liver Disease Activity Score (NAS: steatosis, lobular inflammation, and ballooning) were determined by a different investigator also blind to the clinical status of the patients.

Statistical analysis

To detect statistically significant differences between MH and MUH individuals with an OR=2, level of trust of 0.95, a potency of 0.8, control exposure to the risk factor (systemic inflammation, assessed by hsCRP levels) of 0.2 and case exposure of 0.8, we determined a sample size of 14 patients per group. This sample size is coherent with the sample size

reported in the literature for human studies on adipose tissue NLRP3 inflammasome in obese subjects.³⁰

The results are expressed as total numbers, percentages, and mean \pm SD. For continuous data, statistical significance was determined with non-paired Student's t-test or one-way analysis of variance (ANOVA) test for comparisons of two or three groups, respectively. Tukey post-test was applied after ANOVA to compare specific groups. For non-continuous data, a χ^2 test was performed. Correlation analysis was performed with Pearson's test. All statistical analyses were performed with GraphPad Prism software V.8 (<http://www.graphpad.com/scientific-software/prism/>). Differences were considered significant when the p value was <0.05.

RESULTS

Anthropometric, clinical and laboratory characterization of the patients

Forty-six subjects were included in the study. Based on ATP III criteria for MS and American Diabetes Association (ADA) criteria for T2D diagnosis, individuals were categorized as 'MH' (n=11) and 'MUH' (n=35). The MH group was composed of subjects with zero or one ATP III criterion. Although subjects with two of the ATP III criteria are considered as not having MS in clinical guidelines, they have a greater frequency of dysglycemia, dyslipidemia, and increased waist circumference. Therefore, we excluded this intermediate group (n=14) to extreme the phenotypical differences between MH and MUH individuals.

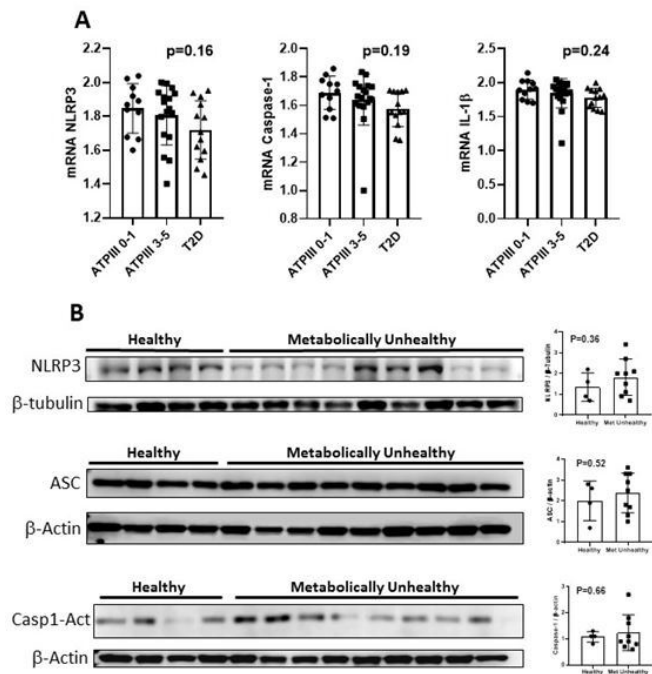


Figure 1 Hepatic mRNA and protein expression of the inflammasome components in healthy and metabolically unhealthy subjects. (A) Hepatic mRNA levels of *Nlrp3*, *Caspase-1*, and *IL-1β*. Each row shows the mean and SD values. Additionally, each row shows single values per group. Statistical analysis performed with one-way analysis of variance. There were no differences in the liver expression of *Nlrp3*, *Caspase-1* or *IL-1β* between groups. Excluding the outlier in group ATP III 3–5 did not change the results. (B) Western blots of *nlrp3*, *asc* and active caspase-1. Graphs of the expression of the western blots of *nlrp3*, *asc* and active caspase-1. Statistical analysis performed with Student t-test. There were no differences between groups in any protein expression. ATP III, Adult Treatment Panel III; IL, interleukin; *Nlrp3*, nod-like receptor protein 3; T2D, type 2 diabetes.

MUH group was composed of patients with MS (n=21) and T2D (n=14). The characteristics of these patients are summarized in [table 1](#). We found no differences in gender and age between MH and MUH patients, although individuals in the MH group tended to be younger and have a higher proportion of women. As expected, Body Mass Index (BMI), waist circumference, and arterial hypertension were significantly higher in MUH subjects. Subgroup analysis showed no differences in these parameters between individuals with MS and individuals with T2D.

As predicted, plasma glucose and insulin levels, homeostatic model assessment of insulin resistance (HOMA-IR), glycated hemoglobin (HbA1c), and hsCRP were significantly higher in MUH as compared to MH individuals ([table 1](#)). Dynamic evaluation of insulin sensitivity showed that MUH individuals had a lower $OGIS_{120}$ index, suggesting higher levels of systemic IR. Subgroup analysis revealed that glycemia and HbA1c were both significantly higher in subjects with T2D as compared to subjects with MS. Interestingly, no differences in total and low-density lipoprotein cholesterol between MUH and MH subjects

were detected, likely as a result of the higher use of statins use in subjects with T2D. High-density lipoprotein cholesterol levels were lower (MH: 54 ± 13 vs MS: 41 ± 11 vs T2D: 42 ± 11 mg/dL, $p=0.006$), and triglycerides levels were higher (MH: 86 ± 26 vs MS: 166 ± 82 vs T2D: 175 ± 102 mg/dL, $p=0.01$) in MUH patients ([table 1](#)).

MUH individuals have increased levels of inflammatory cytokines, but not IL-1β, in circulation

Multiplex analysis of plasma cytokines showed significantly higher levels of adiponectin in MH as compared with MUH subjects, whereas on the contrary, inflammatory cytokines PAI-1, IL-6, and TNF-α were all significantly increased in MUH individuals ([table 2](#)). These findings are concordant with the elevated hsCRP levels in these patients ([table 1](#)) and support the hypothesis that MUH patients had a low-grade chronic systemic inflammatory status, as previously reported.⁸ Unexpectedly, we found that plasma levels of IL-1β were equivalent between MH and MUH subjects (0.68 ± 0.25 vs 0.79 ± 0.35 vs 0.8 ± 0.37 , for healthy subjects, subjects with MS and subjects with T2D, respectively, $p=0.62$), suggesting that either systemic IL-1β formation rates are not increased or the degradation rate is elevated in MUH subjects compared with MH subjects.

Hepatic levels of NLRP3 inflammasome components are not elevated in MUH individuals

Murine models of obesity and diabetes suggest that NLRP3 inflammasome is a key pathogenic player for both hepatic inflammation and IR, by generating increased levels of IL-1β. Therefore, we aimed to assess whether the hepatic abundance of NLRP3 inflammasome components was increased in MUH patients. As shown in [figure 1A](#) and contrary to our expected results, we observed no differences in the hepatic abundance of *Nlrp3*, *Caspase-1*, and *IL-1β* at the mRNA level between MUH and MH patients. Similarly, we found no correlation between hepatic *Nlrp3*, *Caspase-1* and *IL-1β* mRNA levels, and systemic insulin sensitivity markers (online supplemental table 1).

Because the abundance of the NLRP3 inflammasome components are regulated also at the post-transcriptional level, we assessed *nlrp3*, *asc*, and caspase-1 protein levels in whole liver homogenates. As shown in [figure 1B](#), protein levels of *nlrp3* and *asc*, the two main components of the NLRP3 inflammasome, and cleaved caspase-1,^{43 44} a proxy for inflammasome activation, were equivalent between MUH and MH individuals. Importantly, our western blotting analysis revealed great variability in NLRP3 inflammasome components in the liver of the analyzed individuals ([figure 1B](#)), suggesting interindividual differences in the regulation of the levels of NLRP3 inflammasome components.

MUH patients have a more severe hepatic tissue damage that does not correlate with NLRP3 inflammasome levels

Histological analysis showed a higher frequency of normal histology and simple steatosis in MH compared

Table 3 Liver enzymes and histological analysis of healthy controls and subjects with metabolic syndrome and T2D

	Healthy controls	Metabolic syndrome	T2D	P value
	ATP III 0–1	ATP III 3–5	Any ATP III	
	(n=11)	(n=21)	(n=14)	
Liver enzymes				
Aspartate aminotransferase (mg/dL) (mean±SD)	22±6	24±8	28±10	0.14
Alanine aminotransferase (mg/dL) (mean±SD)	19±7	34±20	39±18	0.02*
Liver biopsy				
Normal, n (%)	7 (64)	5 (24)	1 (7)	0.0016†
Steatosis, n (%)	3 (27)	7 (33)	1 (7)	
NASH, n (%)	1 (9)	9 (43)	12 (86)	
Steatosis percentage (mean±SD)	9.1±12.2	26.6±25.5	37.9±28.7	0.01*
Ballooning (mean±SD)	0.09±0.3	0.47±0.6	1.07±0.62	0.0002*‡
Lobular inflammation (mean±SD)	0.27±0.6	0.57±0.6	1.21±0.8	0.003*‡
Fibrosis (mean±SD)	0.09±0.3	0.48±0.6	1.07±0.92	0.002*‡
NAS (mean±SD)	0.82±1.4	2.3±1.9	3.9±2.2	0.001*‡
SAF score (mean±SD)	0.64±1	2.19±1.99	3.79±2.3	0.008*‡

Statistical analysis was performed with one-way analysis of variance and Tukey post-test to compare pairs of groups.

*Denotes significant difference between ATP III 0–1 vs T2D.

†Denotes χ^2 statistical analysis. A significant difference was considered when the p value was <0.05.

‡Denotes significant difference between ATP III 3–5 vs T2D.

ATP III, Adult Treatment Panel III; NAS, Non-alcoholic Fatty Liver Disease Activity Score; NASH, non-alcoholic steatohepatitis; SAF, Steatosis, Inflammatory Activity, and Fibrosis; T2D, type 2 diabetes.

with MUH individuals. On the contrary, MUH patients, either with MS or T2D, had a higher frequency of NASH (table 3). Also, subjects with T2D showed a significantly higher frequency of ballooning, lobular inflammation, and fibrosis as compared with MS subjects. Concordantly, NAS and SAF scores were higher in MUH individuals as compared with MH patients (table 3).

To assess the role of the NLRP3 inflammasome in liver pathology, we performed correlation analysis between the abundance of *Nlrp3*, *Caspase-1*, and *IL-1 β* mRNA with histological markers of liver damage. As shown in figure 2, we found no correlation between the abundance of inflammasome components and NAS and SAF scores as neither with steatosis, lobular inflammation, ballooning, or fibrosis (online supplemental table 1), regardless of the metabolic health status of the patients.

DISCUSSION

Chronic low-grade inflammation correlates and possibly causes hepatic and systemic IR, and the activation of NLRP3 inflammasome is a possible mechanism for hepatic IR and inflammation. Herein, we quantified the hepatic abundance of NLRP3 inflammasome components and the inflammatory status at the systemic and hepatic level in subjects with MS and T2D compared with MH individuals.

Our results show that, as previously reported, MUH subjects have increased circulating levels of several

proinflammatory cytokines as well as hsCRP in comparison with MH individuals, suggesting a state of chronic low-grade systemic inflammation. Also, MUH individuals have a higher frequency of hepatic histopathological indicators of NASH. Nevertheless, in our study, we found no elevations of plasma IL-1 β in MUH patients, suggesting that this cytokine is not implicated in the systemic complications of these patients. Because IL-1 β is the product of NLRP3 inflammasome, we speculate that this inflammasome has no pathophysiological actions in the circulating levels of this cytokine in MUH patients. Nevertheless, it is well possible that inflammasome activation still plays a role in IR and inflammatory processes at specific tissue levels. In fact, it has been reported that increased abundance of NLRP3 inflammasome components in the adipose tissue of obese patients correlates with increased IR in these individuals. However, our results show that the hepatic abundance of NLRP3 inflammasome components at the mRNA and protein levels was not different between MH and MUH subjects, and that NLRP3 inflammasome component abundance did not correlate with static and dynamic indicators of IR (HOMA-IR and OGIS₁₂₀, respectively) nor glycemic control (HbA1c). These results are coherent with our finding that the hepatic abundance of cleaved caspase-1, a surrogate indicator of inflammasome activity,^{43 44} was equivalent between MH and MUH individuals, suggesting that MS and T2D do not increase the abundance nor the

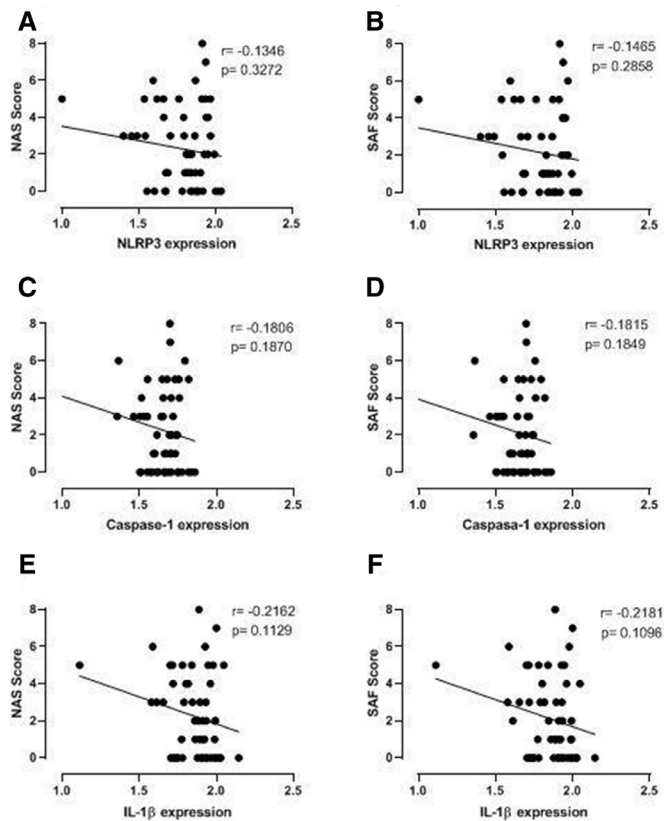


Figure 2 Correlation between mRNA expression of the NLRP3 inflammasome components and NAS and SAF scores. Pearson correlation test did not demonstrate a significant correlation between hepatic mRNA levels of the components of the inflammasome (NLRP3, caspase-1, and IL-1 β) with hepatic NAS and SAF scores. IL, interleukin; NAS, Non-alcoholic Fatty Liver Disease Activity Score; NLRP3, nod-like receptor protein 3; SAF, Steatosis, Inflammatory Activity, and Fibrosis.

activity of hepatic NLRP3 inflammasome. Similarly, we found no correlation between NLRP3 inflammasome component abundance and NAS or SAF scores in the liver, suggesting that the inflammasome is not implicated in the progression of NAFLD. Nevertheless, it must be noted that the small sample size of this study and the lack of direct determinations of inflammasome activity in the liver of the analyzed patients limit the strength of our conclusions.

NLRP3 inflammasome and IL-1 β have been mechanistically linked to IR in murine models of obesity. Mice lacking components of NLRP3 inflammasome are resistant to high-fat diet-induced obesity and IR,²⁵ and the pharmacological inhibition of NLRP3 inflammasome ameliorates the metabolic abnormalities induced by a high-fat diet feeding.⁴⁵ Thus, it is currently proposed that the NLRP3 inflammasome is pathophysiologically linked to T2D by promoting IR as well as by triggering beta-cell dysfunction.^{20 21 24 25}

However, it has been noted that evidence connecting NLRP3 inflammasome and human metabolic diseases

is scarce and indirect, and it is mostly centered in the adipose tissue but not the liver.

The role of NLRP3 inflammasome in human adipose tissue was studied by Esser *et al*, who reported that MUH obese subjects have increased levels of NLRP3 inflammasome components in the visceral adipose tissue compared with MH obese and lean subjects.³⁰ Additionally, these authors observed increased caspase-1 activity and IL-1 β levels in cultured human adipocytes, indirectly supporting a role for this inflammasome in the metabolic complications of obesity. Additionally, Vandanmagsar *et al* described that bodyweight reduction was associated with a reduction in the levels of *Nlrp3* and *IL-1 β* mRNA in the subcutaneous adipose tissue of subjects with T2D, correlating with improved glycemia and insulin sensitivity surrogates.²⁵ On the contrary, Moschen *et al* described that bariatric surgery did not significantly change liver nor subcutaneous adipose tissue *Nlrp3* and *IL-1 β* mRNA levels,⁴⁶ highlighting the controversial roles of the inflammasome in adipose tissue and liver IR.

In HepG2 or L02 cell lines, NLRP3 inflammasome inhibition with molecules such as betaine, liraglutide, Wu Mei Wan and andrographolide, ameliorates IR,⁴⁷⁻⁵⁰ suggesting a role for this inflammasome in hepatic insulin action. However, only studies of primary hepatic diseases such as alcoholic liver disease, liver ischemia/reperfusion, or autoimmune hepatitis⁵¹ have substantiated more direct roles for hepatic NLRP3 inflammasome in human diseases.

In patients with NAFLD and NASH, Wree *et al* reported that hepatic NLRP3 inflammasome components, as well as *Col1A* gene expression, are elevated in patients with extreme obesity (BMI above 50 kg/m²) and NASH compared with patients with simple steatosis.²⁹ However, these authors did not describe the metabolic status of these individuals, which had a much higher BMI and circulating transaminases levels than the patients included in our study. Therefore, although our study does not support an increase in the hepatic levels of NLRP3 components in MUH patients, it remains possible that hepatic NLRP3 inflammasome is increased in severely obese individuals. Nonetheless, it must be noted that even in this subset of patients, the relationship between inflammasome activity, IR, and liver pathology remains formally untested.

In recent years, several post-transcriptional modifications have been described regulating NLRP3 inflammasome activity, including ubiquitination, phosphorylation, nitrosylation, as well as various NLRP3-interacting proteins.^{52 53} Therefore, it is possible that significant changes in inflammasome activity can be present even with unchanged inflammasome component abundance.

In summary, we found that the abundance of hepatic NLRP3 inflammasome components does not correlate with systemic IR and inflammation nor with the histological damage in the liver of MUH patients.

Considering the limitations of this study (small sample size, lack of prospective follow-up, no direct determination of inflammasome activity, and limited ethnical diversity), further studies are required to fully evaluate the role of the hepatic inflammasome system in patients with obesity and its role in IR and NASH.

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Acknowledgements We thank Marco Arrese, MD, for reading this article.

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Funding This study was supported by grants from Fondo Nacional para el Desarrollo Científico y Tecnológico FONDECYT (#11 150 329, awarded to NQ, and #1 181 214, awarded to VC).

Competing interests None declared.

Patient consent for publication Not required.

Ethics approval This study was approved by the ethics committee of the Faculty of Medicine, Pontificia Universidad Católica de Chile, on December 1, 2015 (number of approval 15–313). All patients were included only after they provided their informed consent.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement There are no data in this work.

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REFERENCES

- Besse-Patin A, Estall JL. An intimate relationship between ROS and insulin signalling: implications for antioxidant treatment of fatty liver disease. *Int J Cell Biol* 2014;2014:519153.
- Cani PD, Amar J, Iglesias MA, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* 2007;56:1761–72.
- Houstis N, Rosen ED, Lander ES. Reactive oxygen species have a causal role in multiple forms of insulin resistance. *Nature* 2006;440:944–8.
- Samuel VT, Shulman GI. Nonalcoholic fatty liver disease as a nexus of metabolic and hepatic diseases. *Cell Metab* 2018;27:22–41.
- Eslam M, Newsome PN, Sarin SK, et al. A new definition for metabolic dysfunction-associated fatty liver disease: an international expert consensus statement. *J Hepatol* 2020;73:202–9.
- McArdle MA, Finucane OM, Connaughton RM, et al. Mechanisms of obesity-induced inflammation and insulin resistance: insights into the emerging role of nutritional strategies. *Front Endocrinol* 2013;4:52.
- Saltiel AR, Olefsky JM. Inflammatory mechanisms linking obesity and metabolic disease. *J Clin Invest* 2017;127:1–4.
- Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. *Gastroenterology* 2007;132:2169–80.
- Wieser V, Moschen AR, Tilg H. Inflammation, cytokines and insulin resistance: a clinical perspective. *Arch Immunol Ther Exp* 2013;61:119–25.
- Martinon F, Burns K, Tschopp J. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL- β . *Mol Cell* 2002;10:417–26.
- Latz E. The inflammasomes: mechanisms of activation and function. *Curr Opin Immunol* 2010;22:28–33.
- Man SM, Kanneganti T-D. Regulation of inflammasome activation. *Immunol Rev* 2015;265:6–21.
- Monie TP. NLR activation takes a direct route. *Trends Biochem Sci* 2013;38:131–9.
- Zambetti LP, Laudis F, Licandro G, et al. The rhapsody of NLRPs: master players of inflammation...and a lot more. *Immunol Res* 2012;53:78–90.
- Gao D, Madi M, Ding C, et al. Interleukin-1 β mediates macrophage-induced impairment of insulin signaling in human primary adipocytes. *Am J Physiol Endocrinol Metab* 2014;307:E289–304.
- Jager J, Grémeaux T, Cormont M, et al. Interleukin-1 β -induced insulin resistance in adipocytes through down-regulation of insulin receptor substrate-1 expression. *Endocrinology* 2007;148:241–51.
- Lagathu C, Yvan-Charvet L, Bastard J-P, et al. Long-term treatment with interleukin-1 β induces insulin resistance in murine and human adipocytes. *Diabetologia* 2006;49:2162–73.
- Larsen CM, Faulenbach M, Vaag A, et al. Sustained effects of interleukin-1 receptor antagonist treatment in type 2 diabetes. *Diabetes Care* 2009;32:1663–8.
- Larsen CM, Faulenbach M, Vaag A, et al. Interleukin-1-receptor antagonist in type 2 diabetes mellitus. *N Engl J Med* 2007;356:1517–26.
- Sokolova M, Sahraoui A, Høyem M, et al. NLRP3 inflammasome mediates oxidative stress-induced pancreatic islet dysfunction. *Am J Physiol Endocrinol Metab* 2018;315:E912–23.
- Youm Y-H, Adijiang A, Vandanmagsar B, et al. Elimination of the NLRP3-ASC inflammasome protects against chronic obesity-induced pancreatic damage. *Endocrinology* 2011;152:4039–45.
- Ralston JC, Lyons CL, Kennedy EB, et al. Fatty acids and NLRP3 inflammasome-mediated inflammation in metabolic tissues. *Annu Rev Nutr* 2017;37:77–102.
- Stienstra R, Joosten LAB, Koenen T, et al. The inflammasome-mediated caspase-1 activation controls adipocyte differentiation and insulin sensitivity. *Cell Metab* 2010;12:593–605.
- Stienstra R, Tack CJ, Kanneganti T-D, et al. The inflammasome puts obesity in the danger zone. *Cell Metab* 2012;15:10–18.
- Vandanmagsar B, Youm Y-H, Ravussin A, et al. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat Med* 2011;17:179–88.
- Stienstra R, van Diepen JA, Tack CJ, et al. Inflammasome is a central player in the induction of obesity and insulin resistance. *Proc Natl Acad Sci U S A* 2011;108:15324–9.
- Cai C, Zhu X, Li P, et al. Nlrp3 deletion inhibits the non-alcoholic steatohepatitis development and inflammation in Kupffer cells induced by palmitic acid. *Inflammation* 2017;40:1875–83.
- He K, Zhu X, Liu Y, et al. Inhibition of NLRP3 inflammasome by thioredoxin-interacting protein in mouse Kupffer cells as a regulatory mechanism for non-alcoholic fatty liver disease development. *Oncotarget* 2017;8:37657–72.
- Wree A, McGeough MD, Peña CA, et al. NLRP3 inflammasome activation is required for fibrosis development in NAFLD. *J Mol Med* 2014;92:1069–82.
- Esser N, L'homme L, De Roover A, et al. Obesity phenotype is related to NLRP3 inflammasome activity and immunological profile of visceral adipose tissue. *Diabetologia* 2013;56:2487–97.

- 31 Alberti KGMM, Zimmet P, Shaw J, *et al.* The metabolic syndrome--a new worldwide definition. *Lancet* 2005;366:1059–62.
- 32 National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). Third report of the National cholesterol education program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel III) final report. *Circulation* 2002;106:3143–421.
- 33 Villanueva B, Arteaga A, Maiz A, *et al.* Abdominal obesity is a common finding in normal and overweight subjects of Chile and is associated with increased frequency of cardiometabolic risk factors. *PLoS One* 2018;13:e0194644.
- 34 American Diabetes Association. 2. Classification and Diagnosis of Diabetes: *Standards of Medical Care in Diabetes-2019*. *Diabetes Care* 2019;42:S13–28.
- 35 Mari A, Pacini G, Murphy E, *et al.* A model-based method for assessing insulin sensitivity from the oral glucose tolerance test. *Diabetes Care* 2001;24:539–48.
- 36 Quezada N, León F, Martínez J, *et al.* Emergency right hepatectomy after laparoscopic Tru-cut liver biopsy. *Int J Surg Case Rep* 2015;8C:42–4.
- 37 Khalifa A, Rockey DC. The utility of liver biopsy in 2020. *Curr Opin Gastroenterol* 2020;36:184–91.
- 38 Tapper EB, Lok AS-F. Use of liver imaging and biopsy in clinical practice. *N Engl J Med* 2017;377:756–68.
- 39 Imamura H, Kawasaki S, Bandai Y, *et al.* Comparison between wedge and needle biopsies for evaluating the degree of cirrhosis. *J Hepatol* 1993;17:215–9.
- 40 Guzman-Lepe J, Cervantes-Alvarez E, Collin de l'Hortet A, *et al.* Liver-enriched transcription factor expression relates to chronic hepatic failure in humans. *Hepatol Commun* 2018;2:582–94.
- 41 Valanejad L, Cast A, Wright M, *et al.* PARP1 activation increases expression of modified tumor suppressors and pathways underlying development of aggressive hepatoblastoma. *Commun Biol* 2018;1:67.
- 42 Wadkin JCR, Patten DA, Kamarajah SK, *et al.* CD151 supports VCAM-1-mediated lymphocyte adhesion to liver endothelium and is upregulated in chronic liver disease and hepatocellular carcinoma. *Am J Physiol Gastrointest Liver Physiol* 2017;313:G138–49.
- 43 Martinon F, Tschopp J. Inflammatory caspases and inflammasomes: master switches of inflammation. *Cell Death Differ* 2007;14:10–22.
- 44 Schroder K, Tschopp J. The inflammasomes. *Cell* 2010;140:821–32.
- 45 Chiazza F, Couturier-Maillard A, Benetti E, *et al.* Targeting the NLRP3 inflammasome to reduce diet-induced metabolic abnormalities in mice. *Mol Med* 2016;21:1025–37.
- 46 Moschen AR, Molnar C, Enrich B, *et al.* Adipose and liver expression of interleukin (IL)-1 family members in morbid obesity and effects of weight loss. *Mol Med* 2011;17:840–5.
- 47 Kim DH, Kim SM, Lee B, *et al.* Effect of betaine on hepatic insulin resistance through FOXO1-induced NLRP3 inflammasome. *J Nutr Biochem* 2017;45:104–14.
- 48 Lee HJ, Yeon JE, Ko EJ, *et al.* Peroxisome proliferator-activated receptor-delta agonist ameliorated inflammasome activation in nonalcoholic fatty liver disease. *World J Gastroenterol* 2015;21:12787–99.
- 49 Wan X, Xu C, Lin Y, *et al.* Uric acid regulates hepatic steatosis and insulin resistance through the NLRP3 inflammasome-dependent mechanism. *J Hepatol* 2016;64:925–32.
- 50 Yang X, Li L, Fang K, *et al.* Wu-Mei-Wan reduces insulin resistance via inhibition of NLRP3 inflammasome activation in HepG2 cells. *Evid Based Complement Alternat Med* 2017;2017:1–10.
- 51 Neumann K, Schiller B, Tiegs G. NLRP3 inflammasome and IL-33: novel players in sterile liver inflammation. *Int J Mol Sci* 2018;19. doi:10.3390/ijms19092732. [Epub ahead of print: 12 Sep 2018].
- 52 Kelley N, Jeltema D, Duan Y, *et al.* The NLRP3 inflammasome: an overview of mechanisms of activation and regulation. *Int J Mol Sci* 2019;20. doi:10.3390/ijms20133328. [Epub ahead of print: 06 Jul 2019].
- 53 Yabal M, Calleja DJ, Simpson DS, *et al.* Stressing out the mitochondria: mechanistic insights into NLRP3 inflammasome activation. *J Leukoc Biol* 2019;105:377–99.