

Cord serum metabolome and birth weight in patients with gestational diabetes treated with metformin, insulin, or diet alone

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ABSTRACT

Introduction Recent research has demonstrated the benefits of metformin treatment in gestational diabetes (GDM) on short-term pregnancy outcomes (including excessive fetal growth and pre-eclampsia), but its effects on fetal metabolism remain mostly unknown. Our aim was to study the effects of metformin treatment compared with insulin or diet on the cord serum metabolome and also to assess how these metabolites are related to birth weight (BW) in pregnancies complicated by GDM.

Research design and methods Cord serum samples were available from 113, 97, and 98 patients with GDM treated with diet, insulin, and metformin, respectively. A targeted metabolome was measured using nuclear magnetic resonance spectroscopy. The patients in the metformin and insulin groups had participated in a previous randomized trial (NCT01240785).

Results Cord serum alanine was elevated in the metformin group (0.53 mmol/L) compared with the insulin (0.45 mmol/L, $p < 0.001$) and the diet groups (0.46 mmol/L, $p < 0.0001$). All other measured metabolites were similar between the groups. The triglyceride (TG)-to-phosphoglyceride ratio, average very low-density lipoprotein particle diameter, docosahexaenoic acid, omega-3 fatty acids (FAs), and ratios of omega-3 and monounsaturated FA to total FA were inversely related to BW. The omega-6-to-total-FA and omega-6-to-omega-3-FA ratios were positively related to BW. Cholesterol in very large and large high-density lipoprotein (HDL) was positively ($p < 0.01$) associated with BW when adjusted for maternal prepregnancy body mass index, gestational weight gain, glycated hemoglobin, and mode of delivery.

Conclusions Metformin treatment in GDM leads to an increase in cord serum alanine. The possible long-term implications of elevated neonatal alanine in this context need to be evaluated in future studies. Although previous studies have shown that metformin increased maternal TG levels, the cord serum TG levels were not affected. Cord serum HDL cholesterol and several FA variables are related to the regulation of fetal growth in GDM. Moreover, these associations seem to be independent of maternal confounding factors.

Trial registration number NCT01240785.

INTRODUCTION

Gestational diabetes (GDM) is a growing health concern that affects both mothers

Significance of this study

What is already known about this subject?

- Metformin causes alterations in the maternal metabolome during pregnancy.
- Metformin may be superior to insulin regarding short-term perinatal outcomes. However, the short-term and long-term metabolic implications for the offspring are not known.

What are the new findings?

- The offspring of metformin-treated mothers had higher cord serum alanine levels compared with the insulin-treated and diet-treated groups.
- Metformin treatment in gestational diabetes does not appear to affect the cord serum lipid profile or cord serum lactate or ketone levels.

How might these results change the focus of research or clinical practice?

- Although metformin causes an elevation in maternal triglycerides, it was reassuring to find that the lipid profile of the neonate as examined using cord serum was not affected.
- The long-term implications of elevated cord serum alanine need to be evaluated in the future.

and neonates worldwide. It increases the risk of complications such as macrosomia, pre-eclampsia, neonatal hypoglycemia and hyperbilirubinemia, and the need for neonatal intensive care.¹ In the long term, GDM increases especially the risk for type 2 diabetes in mothers² and also metabolic syndrome and obesity in the offspring.³ The most common pharmacological treatment options for GDM are insulin and metformin; both affect maternal glucose, lipid,⁴ and amino acid⁵ metabolism.

When diet and lifestyle modifications alone are insufficient to control maternal hyperglycemia, a pharmacological treatment (either metformin or insulin) is initiated. Compared with insulin, metformin appears superior at reducing risk of neonatal hypoglycemia,

macrosomia, large-for-gestational age (LGA) neonate, pre-eclampsia, and neonatal intensive care unit admission.⁶ However, metformin crosses the placenta⁷; in follow-up studies, metformin exposure in utero has been associated with increased prepubertal anthropometric variables such as weight, Body Mass Index (BMI), and waist circumference in offspring,^{8,9} which has led to concerns regarding metformin's long-term effects on offspring's health. Whether these effects are promoted by alterations in fetal metabolism remains unknown. A previous trial comparing metformin and insulin treatment in GDM revealed no differences in cord serum triglyceride (TG), high-density lipoprotein (HDL) cholesterol, and low-density lipoprotein (LDL) cholesterol levels.¹⁰ To our knowledge, no previous studies have evaluated the effect of antenatal metformin exposure on the cord serum metabolome (a proxy of fetal metabolism) in detail.

In addition to genetic factors, adequate nutrition, and placental blood flow, ample evidence supports the role of maternal serum metabolites in fetal growth.¹¹ Particularly in pregnancies complicated by GDM, maternal serum glucose,¹ lipids,¹² and amino acids^{5,13} have been related to fetal growth. Furthermore, it has been shown that cord serum metabolites are associated with birth weight (BW)^{14,15} and may also be associated with the long-term metabolic health of the offspring, although these studies have yielded divergent results.^{16,17}

The aim of this study was to evaluate the possible effects of metformin treatment of GDM in the neonate cord serum targeted metabolome compared with maternal treatment with insulin or lifestyle modifications alone. Our second aim was to evaluate the clinical impact of the variation in cord serum metabolome. Due to low overall incidence of adverse perinatal outcomes in the study¹⁸ and the apparent dysregulation of fetal growth in GDM, BW was chosen as the outcome of interest. Thus, we examined to which extent these metabolites are associated with BW in GDM and whether these associations are modified by GDM treatment.

RESEARCH DESIGN AND METHODS

The study population treated with antihyperglycemic drugs was derived from a previous randomized trial that compared metformin and insulin in participants with GDM.¹⁸ In addition, patients diagnosed with GDM but not requiring pharmacological treatment (described in¹⁹) were included in the present study. The study participants were recruited at Turku University Hospital (Finland) between June 2006 and December 2010. The GDM diagnosis was made according to Finnish national criteria based on a 2-hour 75 g oral glucose tolerance test (OGTT). The diagnostic cut-off values were ≥ 4.8 (fasting), ≥ 10.0 (1 hour) and ≥ 8.7 mmol/L (2 hours) until the release of new guidelines in December 2008; thereafter, cut-off values were ≥ 5.3 , ≥ 10.0 , and ≥ 8.6 mmol/L, respectively. At least two elevated values were required. The patients

in the study were diagnosed at a mean of 27.0 (SD=2.6) gestational weeks. Pharmacological treatment was indicated if the patient had recurrent hyperglycemia (fasting glucose ≥ 5.5 and/or postprandial glucose ≥ 7.8 mmol/L) despite diet and lifestyle modifications. The patients who required pharmacological treatment were randomized to either the metformin or insulin groups at a mean of 30 gestational weeks, as described previously.¹⁸ Metformin was started at a daily dose of 500 mg and increased up to 2000 mg if needed (median: 1500 mg). For insulin treatment, neutral protamine Hagedorn insulin and/or rapid-acting insulin (lispro or aspart) were used. The study was registered with ClinicalTrials.gov (<http://clinicaltrials.gov/ct2/show/NCT01240785>).

Glycated hemoglobin (HbA1c) and fasting serum C-peptide levels were analyzed at the time of the GDM diagnosis using routine laboratory methods. BW was expressed in gram and SD units (the deviation from the Finnish population mean BW adjusted for gestation duration).²⁰ Small-for-gestational age (SGA) and LGA neonates were classified using BWs >90 th percentile and <10 th percentile, respectively.

In the present study, we had cord serum samples available from 113, 97, and 98 neonates in the diet, insulin, and metformin groups, respectively. Twenty-one percent of patients who were randomized to receive metformin required additional insulin. These patients were included in the metformin group in the analyses because we argued that the possible untoward metabolic effects of metformin exposure would also be present in patients receiving both metformin and insulin. Moreover, these patients were older and had higher HbA1c at the time of diagnosis than the rest of the metformin group, and therefore exclusion of these patients would introduce bias. The effect of exclusion was, however, evaluated in a sensitivity analysis. Mixed arterial and venous cord blood was taken immediately after delivery from the placental side of the cut umbilical cord by letting the blood flow freely into test tubes. The samples were allowed to clot at room temperature and then were centrifuged at 3000 rpm for 5 min to obtain serum, which was stored at -70°C prior to further analyses.

A targeted metabolic biomarker profile that included lipids, lipoproteins, amino acids, glycolysis-related metabolites, and ketones (142 metabolic measures in total, online supplemental table 1) was analyzed using high-throughput proton nuclear magnetic resonance (NMR) spectroscopy (Nightingale Health, Helsinki, Finland).²¹ The sample preparation, NMR spectra acquisition, metabolite identification, and quantification process have been described in detail in the online supplemental file 1). Due to the rejection of individual measurements by quality control, there was a slight difference in the number of successfully quantified samples for various metabolic measures. The NMR protocol has been widely applied in epidemiological research, and the quantification accuracy has shown high consistency when compared with routine clinical chemistry methods (eg,

the correlation coefficients between NMR and routine clinical chemistry methods ranged from 0.91 to 0.96 for glucose, TG, and total, HDL and LDL cholesterol).²²

Statistical analysis

The differences in the baseline characteristics of the mothers and the neonates were evaluated using an analysis of variance and Tukey's honestly significant difference test or the Kruskal-Wallis test and the Dwass-Steele test, depending on the distributions. A comparison of categorical data was carried out using the χ^2 test or Fisher's exact test. The medians of individual metabolites were compared between the groups using the Kruskal-Wallis and the Dwass-Steele tests. For each metabolite, the association with BW was assessed by the following: (1) an unadjusted linear regression analysis; (2) a linear regression analysis adjusted for possible confounding factors determined a priori (mode of delivery, maternal prepregnancy Body Mass Index (pBMI), gestational weight gain (GWG), and HbA1c); and (3) if there was a significant association in the previous regression analyses, a possible interaction between metabolite and treatment group was evaluated both with and without additional confounding factors. Mode of delivery acts as a surrogate of fetal stress during the labor, which may affect cord serum metabolites, and was thus included as covariant in the analyses.

Statistical analyses were completed using R statistical software V.3.6.1 (<http://cran.r-project.org>). To limit the possibility of type I error, a p value of <0.01 was considered statistically significant in the metabolite analyses. Otherwise, for interaction terms and differences in maternal and neonatal characteristics, a p value threshold of 0.05 was employed.

RESULTS

Population characteristics are shown in [table 1](#). The OGTT 1-hour glucose value was lower in the diet group compared with the metformin group (10.9 ± 1.05 vs 11.3 ± 1.47 mmol/L, $p=0.0061$), and the incidence of induction of labor was higher in the insulin group (55.7%) compared with the diet (31.9%, $p=0.0025$) or the metformin group (36.7%, $p=0.036$). Maternal age, HbA1c at the time of diagnosis, and OGTT fasting and 2-hour glucose values tended to be lower in the diet group, although none of these differences were significant according to between-group comparisons. Otherwise, the three groups were similar regarding maternal and neonatal baseline characteristics and clinical outcome data.

Cord serum alanine was significantly higher in the offspring of metformin-treated mothers (0.53 mmol/L) compared with the insulin (0.45 mmol/L, $p<0.001$) or the diet groups (0.46 mmol/L; $p<0.0001$; [figure 1](#)). No other metabolite was significantly different between the three groups (online supplemental table 1). Within the metformin group, cord serum metabolites were similar between all pregnancies with or without additional insulin

treatment. Moreover, the cord serum alanine remained higher in the metformin group compared with other groups even after excluding the pregnancies treated with additional insulin (online supplemental table 2).

The associations between cord serum metabolites and BW were first evaluated using unadjusted regression analyses ([figure 2](#)). Very low-density lipoprotein (VLDL) particle size was inversely related to BW (-0.26 ; 95% CI -0.37 to -0.15 SD/SD, $p<0.0001$), as was the ratio of TG to phosphoglycerides (-0.24 ; 95% CI -0.36 to -0.13 SD/SD, $p<0.0001$). Omega-3 fatty acid (FA) levels, omega-3-to-total-FA ratio, and the docosahexaenoic acid (DHA) and monounsaturated fatty acid (MUFA)-to-total-FA ratio were inversely related to BW. The omega-6-to-total-FA ratio, the omega-6-to-omega-3-FA ratio and the MUFA-to-polyunsaturated-fatty acid (PUFA) ratio were positively associated with BW.

After adjusting for pBMI, GWG, maternal HbA1c, and the mode of delivery cholesterol level in very large and large HDL, total lipids in very large HDL, and average HDL particle size were significantly associated with BW ([figure 2](#)). The ratio of PUFA to total FA was positively and the degree of unsaturation inversely associated with BW when adjusted for the confounding factors.

Histidine was the only amino acid that was significantly and positively associated with BW. Of the glucose metabolism-related metabolites, only 3-hydroxybutyrate and acetone were found to be positively associated with BW.

The association between the omega-6-to-omega-3-FA ratio and BW was the only significant association affected by maternal treatment (interaction term $p<0.05$). This positive association was significant in the insulin group (0.34; 95% CI 0.12, 0.56 SD/SD, $p=0.002$) but not in the metformin (0.22; 95% CI -0.010 to 0.45 SD/SD, $p=0.06$) or the diet group (0.045; 95% CI -0.12 to 0.21 SD/SD, $p=0.59$). After adjusting for confounding factors, the interaction terms were no longer significant.

DISCUSSION

In the present study, we found that compared with insulin or diet treatment, metformin caused a similar rise in cord serum alanine as seen in the serum of women with GDM⁵ and in non-pregnant subjects compared with placebo.²³ Furthermore, a large VLDL particle size, a high TG-to-phosphoglycerides ratio, elevated MUFA-to-total-FA ratio, increased DHA level, and increased omega-3 FA level were associated with a smaller BW, whereas the proportion of omega-6 FA of the total FA was associated with a higher BW.

Elevated cord blood alanine has been found in the presence of GDM.^{24,25} However, to our knowledge, the effects of metformin treatment on neonatal serum alanine have not been studied previously. In line with previous findings on the maternal serum amino acid profile⁵ and in non-pregnant individuals,^{23,26} the cord serum alanine concentration was significantly higher in the metformin

Table 1 Study population characteristics

Variable	Diet	n	Insulin	n	Metformin	n	P value
Maternal characteristics							
Age (years)	30.3±5.2	113	31.8±5.5	97	32.1±5.1	98	0.026 a
Smoking	11 (9.82)	112	16 (16.7)	96	9 (9.57)	94	0.25
Primipara	52 (46)	113	45 (46.4)	97	37 (37.8)	98	0.38
Prepregnancy BMI (kg/m ²)	29.1±5.5	110	28.8±4.7	97	29.6±6.1	98	0.59
HbA1c at GDM diagnosis (%)	5.38±0.33	113	5.50±0.35	97	5.48±0.34	98	0.02 b
OGTT fasting glucose (mmol/L)	5.38±0.42	113	5.54±0.42	97	5.52±0.55	98	0.029 b
OGTT 1-hour glucose (mmol/L)	10.9±1.0	113	11.2±1.2	97	11.3±1.5	98	0.0099 aa
OGTT 2-hour glucose (mmol/L)	7.89±1.87	113	7.84±1.74	96	8.32±1.78	97	0.042
Fasting C-peptide at GDM diagnosis (nmol/L)	1.01±0.31	95	1.05±0.30	91	1.06±0.33	93	0.41
Pregnancy outcomes							
Gestational hypertension	4 (3.54)	113	3 (3.09)	97	2 (2.04)	98	0.84
Pre-eclampsia	4 (3.54)	113	9 (9.28)	97	5 (5.1)	98	0.22
Operative vaginal delivery	7 (6.19)	113	7 (7.22)	97	9 (9.18)	98	0.71
Cesarean delivery	17 (15)	113	15 (15.5)	97	14 (14.3)	98	0.97
Induction of labor	36 (31.9)	113	54 (55.7)	97	36 (36.7)	98	0.0013 bbb, c
Gestational weight gain (kg)	8.6±5.1	111	7.8±5.3	97	8.0±5.3	98	0.47
Gestational age at delivery (weeks)	39.3±2.2	113	39.3±1.6	97	39.2±1.4	98	0.83
Neonatal outcomes							
BW (g)	3530±550	113	3560±440	97	3580±500	98	0.78
BW (SD)	-0.08±1.06	113	0.12±0.93	97	0.14±1.08	95	0.2
BW (centiles)	0.47±0.29	113	0.54±0.29	97	0.54±0.29	95	0.18
SGA	15 (13.3)	113	8 (8.25)	97	12 (12.6)	95	0.46
LGA	12 (10.6)	113	15 (15.5)	97	14 (14.7)	95	0.53
Admission to NICU	33 (29.2)	113	34 (35.1)	97	30 (30.6)	98	0.64
Neonate intravenous glucose	26 (23.4)	111	21 (21.9)	96	23 (23.5)	98	0.96

Data are given as mean±SD or n (%).

P value is given for comparison of the whole population, and significant differences between each pair of groups are denoted with lower-case letters: a, diet–metformin $p<0.05$; aa, diet–metformin $p<0.01$; b: diet–insulin $p<0.05$; bbb: diet–insulin $p<0.001$, c, insulin–metformin $p<0.05$.

LGA: BW>90th centile; SGA: BW<10th centile.

BMI, Body Mass Index; BW, birth weight; GDM, gestational diabetes; HbA1c, glycated hemoglobin; LGA, large for gestational age; NICU, neonatal intensive care unit; OGTT, oral glucose tolerance test; SGA, small for gestational age.

group compared with the insulin or diet groups in this study. This finding could be the result of altered placental metabolism or transfer, increased placental transfer in the presence of an elevated maternal gradient, or due to changes in fetal metabolism. Research in both sheep and human pregnancies has demonstrated that the rate of direct alanine transfer from maternal to fetal circulation is minimal, compared with the effects of placental metabolism and placental–fetal transfer.^{27 28} Fetal alanine concentrations are, however, highly influenced by maternal concentrations.^{27 28} Metformin crosses the placenta, and concentrations similar to maternal serum metformin are present in fetal blood.⁷ Thus, by inhibiting hepatic gluconeogenesis,²⁹ metformin could have

caused an increase in fetal alanine and other gluconeogenic substrates, that is, lactate, alike seen in maternal serum.⁵ Although gluconeogenesis in the human fetus is not currently well characterized, the lack of differences found in our study in other gluconeogenic substrates (pyruvate, lactate, glycerol, and glutamine) or in cord serum glucose reduces the likelihood of major alterations in fetal metabolism in this population due to maternal GDM treatment. Therefore, it seems more plausible that the elevated alanine level in the cord serum was due to increased placental release of alanine rather than altered fetal alanine metabolism.

Metformin treatment of GDM has been associated with a higher weight of the offspring during childhood.⁹

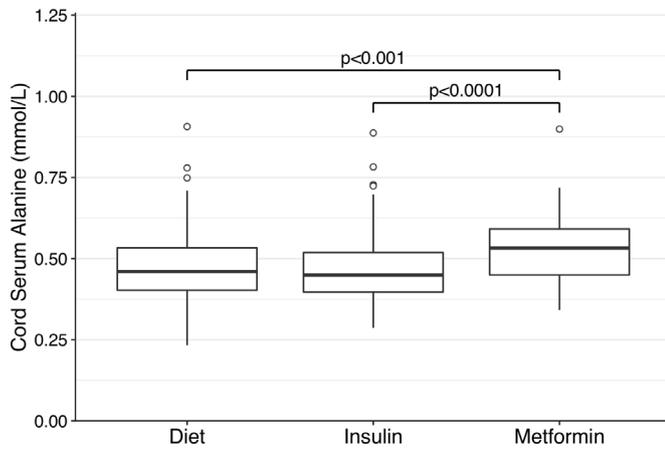


Figure 1 Cord serum alanine concentrations in the diet-treated, insulin-treated, and metformin-treated groups. Boxplot representation of the distributions of cord serum alanine concentrations in the diet-treated, insulin-treated and metformin-treated groups, respectively. IQRs with medians are marked with boxes, and the whiskers denote the range. Outliers, which were defined as being more than 1.5 times the IQR away from the median, are marked with circles. P values are given for differences in the Dwass-Steele test and only significant values are shown.

In a birth cohort study, cord serum alanine was positively related to postnatal weight gain and BMI at 2 and 15 years of age, but the association was not significant after Bonferroni correction.¹⁴ Whether increased fetal serum alanine during late pregnancy could be related to an increased risk of weight gain later in postnatal life warrants further studies.

The degree of maternal glycemia is strongly related with BW,¹ and measures of maternal glycemia (HbA1c) and endogenous insulin production (C-peptide) have previously been shown to be correlated with both alanine⁵ and BW.¹⁰ Although alanine concentration in the maternal serum was related to BW,⁵ in the present study, alanine concentration in the cord serum was not. Therefore, higher maternal alanine levels may reflect maternal insulin resistance, and insulin resistance seems to be the common denominator explaining the association between maternal alanine and BW. Consequently, the ability of alanine to accelerate fetal growth directly seems less plausible.

In fetal sheep, the infusion of alanine resulted in elevations of glucose and insulin secretion.³⁰ The achieved alanine concentrations were 0.50–0.75 mmol/L higher compared with baseline. In our study, the cord serum alanine level was <0.1 mmol/L higher in the metformin group; therefore, the results of these two studies are not well comparable. Moreover, the cord serum glucose was not higher in neonates exposed to metformin compared with the insulin and diet groups. Previously in a birth cohort study, the cord vein alanine level had an inverse association with BW, although this finding was not significant after Bonferroni correction.¹⁴ In contrast, in SGA neonates, the cord alanine was decreased;³¹ in another

large study, several cord blood amino acids, but not alanine, were related to BW.³²

Metformin treatment in GDM leads to higher maternal concentrations of total TG,^{4 10} VLDL TG, VLDL cholesterol, MUFA, SFA, and total FA.⁴ However, these changes were not reflected in the cord serum lipid levels.

It has been proposed that in the context of GDM, cord blood TG levels are inversely related to BW, due to a greater amount of adipose tissue in large fetuses and thus enhanced adipose tissue FA uptake from circulating TG.¹² This uptake may be further promoted by fetal hyperinsulinemia in pregnancies complicated by hyperglycemia.¹ Accordingly, in growth-restricted fetuses with presumably less adipose tissue, the cord plasma TG level is elevated.³³ In addition, we also observed an inverse relationship between BW, VLDL particle size, and the TG-to-phosphoglycerides ratio, which both reflect the amount of TG in lipoproteins. In previous studies, the cord vein serum VLDL particle concentration and VLDL TG were elevated in growth-restricted^{33 34} and SGA³⁵ neonates, whereas there was no difference in HDL TG level.³⁴ As the inverse association with BW was most evident in the VLDL lipids that supply peripheral tissues, our results support the hypothesis of increased FA uptake from circulating TG in large fetuses.

While the concentrations of LDL and VLDL in cord serum are low, the HDL concentration is similar to non-pregnant adult serum. The apolipoprotein composition of fetal HDL, however, differs from that of adult serum with a relative abundance of apoE.³⁶ ApoE serves as a ligand for a variety of receptors (including LDL receptors) and therefore the greater ability of fetal HDL to transport cholesterol between fetal tissues. Cord serum HDL cholesterol has been shown to be related to insulin-like growth factor-1 and BW³⁵ and is decreased in SGA neonates,³⁷ although some studies have found no difference between SGA and appropriately grown neonates.³⁴ Thus, it was not unexpected to find a positive association between cord serum cholesterol in very large and large HDL and BW. This association was significant after adjustment for maternal pBMI, GWG, and HbA1c, which suggests that fetal HDL may play a role in fetal growth that is partially independent of maternal characteristics and glycemic control. Moreover, the association was still identified independently of the treatment group.

The importance of PUFA in fetal development has been previously characterized. Long-chain PUFAs, in particular, are required by the central nervous system and for retinal development; during the third trimester, the placenta has selectivity towards the transfer of these lipids. In contrast, maternal omega-6 FA levels have been positively related to infant obesity.³⁸ Similarly, a high omega-6-to-omega-3 ratio in the maternal diet, maternal plasma, and cord plasma are associated with an increased risk of infant obesity at 3 years of age.³⁹ Arachidonic acid, an omega-6 PUFA, may promote differentiation from preadipocytes into adipocytes,⁴⁰ while DHA inhibits this differentiation.⁴¹ Accordingly, we found that high

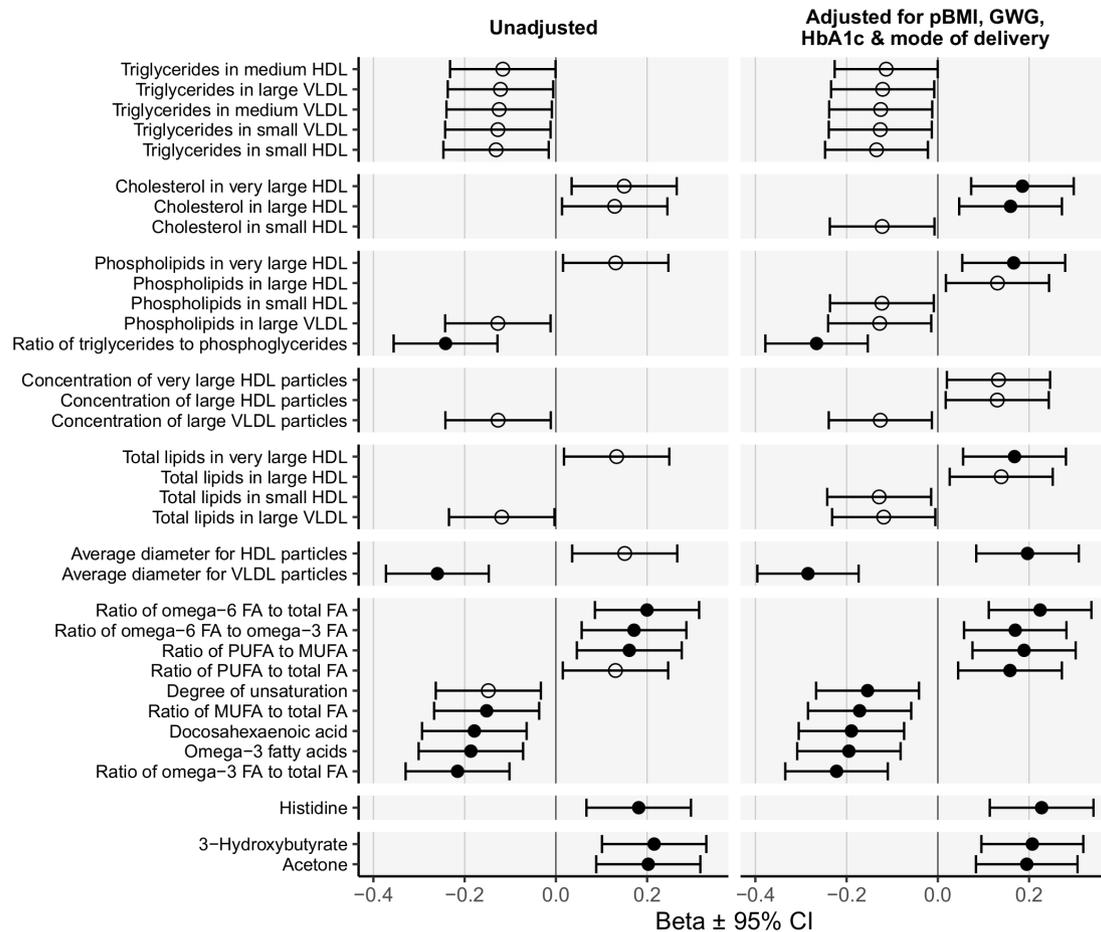


Figure 2 Summary of significant associations between cord serum metabolites and adjusted birth weight. Associations between cord serum metabolites and birth weight in the metformin-treated, insulin-treated, and diet-treated groups combined. Beta values (SD/SD) with 95% CIs are shown for unadjusted linear regressions, and regression models adjusted for maternal BMI, GWG, maternal HbA1c, and mode of delivery. BW was expressed in SD units (ie, deviation from the Finnish population's mean BW adjusted for gestation duration). Open circles denote $p < 0.05$, and closed circles indicate $p < 0.01$. Only associations with $p < 0.05$ are shown. BW, birth weight; FA, fatty acid; GWG, gestational weight gain; HbA1c, glycated hemoglobin; HDL, high-density lipoprotein; MUFA, monounsaturated fatty acid; pBMI, prepregnancy Body Mass Index; PUFA, polyunsaturated fatty acid; VLDL, very low-density lipoprotein.

omega-6-to-omega-3-FA, and omega-6-to-total-FA ratios as well as high PUFA-to-MUFA, and high PUFA-to-total-FA ratios were positively related to BW. Conversely, a high cord serum degree of FA unsaturation and high concentrations of omega-3 FA (including DHA and high omega-3-FA-to-total-FA, and MUFA-to-total-FA ratios) were inversely associated with BW.

Previously, cord blood DHA has been inversely^{42,43} and arachidonic acid has been positively⁴⁴ related to BW, although in some studies, arachidonic acid was also inversely related to BW.⁴³ The divergent results in these two studies and our study may reflect differences in methodology but also the fact that our cord blood samples obtained from neonates exposed to GDM are not representative of the general maternal population. In pregnancies complicated by GDM reduced umbilical arterial omega-3 FA, omega-6 FA, and PUFA levels have been found, suggesting an altered fetal metabolism of these lipids.⁴⁵

The proportion of serum omega-3 and omega-6 FA of total FA decreased in both metformin-treated and insulin-treated patients with GDM during the last half of pregnancy.⁴ However, compared with insulin, metformin led to a greater decrease in the proportion of omega-6 FA and PUFA to total FA. Controversially, omega-6 FA concentration in the maternal serum was associated with BW only in metformin-treated patients,⁴ whereas the association between the cord serum omega-6-to-omega-3-FA ratio and BW was significant only in the insulin-treated group. Because there was no interaction between the treatment group and other associations of BW with cord serum PUFA and omega FA and the regression beta values with 95% CIs were rather similar between the insulin-treated (0.34; 95% CI 0.12 to 0.56 SD/SD) and metformin-treated groups (0.22; 95% CI -0.010 to 0.45 SD/SD), this could be an incidental finding. Moreover, the treatment×metabolite interaction was no longer significant after adjusting for confounding factors.

Our results suggest that a high cord serum omega-6-to-omega-3-FA ratio, which has previously been associated with cardiovascular risk,⁴⁶ is positively associated with BW in the offspring of mothers with GDM. Whether increased fetal adiposity causes increased uptake of MUFA and omega-6 FA or whether the increased omega-6-to-omega-3-FA ratio promotes fetal adipogenesis is not known. Based on previous studies, maternal serum concentrations explain only a small amount of variation in cord serum PUFA,⁴⁷ and although maternal PUFA and omega-6 FA were positively related with BW in metformin-treated patients with GDM,⁴ these associations between cord lipids and BW are not affected by maternal treatment. Because similar associations have been found between maternal diet and the plasma omega-6-to-omega-3-FA ratio and measures of infant obesity,³⁹ future trials of GDM treatment should perhaps focus more specifically on dietary fats.

Histidine was the only amino acid associated with BW. In a previous study, histidine, glycine, and taurine were downregulated in SGA fetuses compared with appropriately growing controls.⁴⁸ In a different population cohort, histidine was positively related to BW.¹⁴ The fact that only cord serum histidine showed an association with BW in our study seems paradoxical since the placental transfer of several other amino acids is also disturbed in GDM.²⁴

Although maternal serum lactate is more elevated in metformin-treated compared with insulin-treated patients with GDM,⁵ there was no difference in cord serum lactate levels. In addition, there was no difference between treatment groups in cord serum ketone levels. We are not aware of any other studies that have reported the effects of intrauterine metformin exposure on cord serum lactate or ketone levels.

In our study population of pregnancies with GDM and good glycemic control, cord serum glucose or lactate levels were not significantly associated with BW. Two ketones (3-hydroxybutyrate and acetone) were positively related with BW. Similarly, Lowe *et al.*¹⁵ suggested a link between cord plasma 3-hydroxybutyrate levels and increased BW and fetal adiposity. Since the fetal liver does not produce a significant amount of ketones but 3-hydroxybutyrate crosses the placenta, the ketones present in cord blood are likely of maternal origin.⁴⁹ In the fetus, ketones may be used for oxidation or lipogenesis, but to which extent they are able to independently promote fetal growth is not known.

Despite the efficacy of metformin treatment in GDM,⁶ prenatal metformin exposure due to placental transfer⁷ has raised some concerns over the possible long-term effects of this treatment. It was thus reassuring to find that the cord serum lipid profile in the metformin group did not significantly differ from either the insulin-treated or the diet-treated group. Moreover, there were no differences in cord serum lactate or ketone levels, suggesting that metformin treatment does not compromise fetal glucose metabolism.

Future studies should focus on whether elevated alanine in the offspring of metformin treated mothers continues during the neonatal period and whether cord serum alanine is associated with long-term metabolic traits. Our study was unable to differentiate whether changes in cord serum alanine were caused by alterations in maternal or fetal metabolism. Future research including simultaneous maternal and fetal serum samples may be better equipped to answer this question.

Strengths and limitations of the study

Our study population represents patients with GDM of mostly Caucasian origin with good glycemic control. The homogeneity of the population simplified the interpretation of the results but also diminished the results' generalizability to other GDM populations. Although the associations between cord serum metabolites and BW were essentially similar between the treatment groups, these associations may be different in pregnancies not complicated by GDM.⁵⁰ We used mixed cord blood samples instead of arterial or venous samples. Although the differences in amino acid, FA, and cholesterol concentrations between umbilical arterial and venous samples are mostly small,^{24 45 51} the proportions of arterial and venous blood may have varied to some degree in individual mixed cord blood samples, thus being a potential source of variation in metabolite concentrations. The sampling method, however, was similar across the study groups and therefore should not have caused bias in the results.

CONCLUSIONS

Metformin treatment in GDM leads to an increase in cord serum alanine but does not affect neonate lipid profile, although metformin has previously been shown to increase maternal TG.^{4 10} Possible long-term effects of increased cord serum alanine in the offspring require further follow-up studies. Our results support the hypothesis of increased FA uptake from circulating TG in larger fetuses. Cord serum cholesterol in very large HDL, the proportions of PUFA and MUFA of all FA, the PUFA-to-MUFA ratio, and the omega-6-to-omega-3-FA ratio are associated with the regulation of fetal growth in GDM. Moreover, these associations seem to be independent of maternal pBMI, GWG, HbA1c, and type of GDM treatment.

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Contributors KT provided clinical data on the metformin-treated and insulin-treated patients and the cord serum samples of all patients from a previous study. OP provided clinical data of the diet treated patients and reviewed the manuscript. MSH analyzed the data and wrote the first draft of the manuscript. KT and TR designed the present study and revised the manuscript. All authors approved the final version of the manuscript.

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Patient consent for publication Not required.

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Data availability statement The data used and/or analyzed during the current study are available from the corresponding author on a reasonable request.

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Supplementary description of the NMR protocol

As described previously, [1,2] the frozen samples were slowly thawed at +4°C and mixed at a ratio of 1:1 with a sodium phosphate buffer (75 mM Na₂HPO₄ in 80%/20% H₂O/D₂O at a pH of 7.4 and also including 0.08% sodium 3-[trimethylsilyl]propionate-2,2,3,3-d₄ and 0.04% sodium azide). Each 96-well plate contained two quality control samples. The prepared samples were warmed to a stabilized temperature of 36.95°C just before the measurements were taken.

The NMR spectra were measured using a Bruker AVANCE III 500-Mhz spectrometer. The spectral data was acquired in two separate parts: a presaturated proton NMR spectrum, which features resonances arising mainly from proteins and lipids within various lipoprotein particles, and a T₂-filtered spectrum where most of the broad macromolecule and lipoprotein lipid signals are suppressed, leading to enhanced detection of low-molecular-weight metabolites. [3] The first spectrum was recorded with 80k data points and 8 scans using Bruker noesyprsat pulse sequence. The second spectrum was recorded with 64k data points and 24 or 16 scans using T₂-relaxation filtered Bruker 1D CPMG pulse sequence. The characteristic molecular spectrums of the spectral windows are presented in [3].

The NMR spectra then underwent Fourier transformation, phase correction, overall signal check for missing/extra peaks, background control, baseline removal, and spectral area-specific signal alignments. [2] The quality control procedures included comparisons with the control samples and an extensive database of quantitative molecular data. Internal standards were not used, but the concentrations were calibrated to comply with external standards. [2] The quantification of metabolites is based on Bayesian modelling. [4,5] The lipid

quantification is calibrated against gel permeation high-performance liquid chromatography as briefly described in, [6] and the consistency with alternative approaches such as mass spectrometry and gas chromatography have been demonstrated. [7,8]

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Supplementary table 1 – Comparison of cord serum metabolites between metformin, insulin, and diet treatment groups

	Unit	Diet		Insulin		Metformin		p-values			
		median [IQR]	n	median [IQR]	n	median [IQR]	n	Overall	Diet vs insulin	Diet vs metformin	Insulin vs metformin
Triglycerides											
Total triglycerides	mmol/l	0.36 [0.28–0.46]	113	0.31 [0.26–0.44]	97	0.32 [0.25–0.43]	98	0.22	0.24	0.39	0.96
Triglycerides in VLDL	mmol/l	0.14 [0.1–0.22]	113	0.12 [0.086–0.21]	97	0.13 [0.076–0.22]	98	0.27	0.28	0.43	0.99
Triglycerides in LDL	mmol/l	0.085 [0.073–0.095]	113	0.077 [0.068–0.089]	97	0.079 [0.068–0.086]	98	0.091	0.17	0.13	1
Triglycerides in HDL	mmol/l	0.057 [0.048–0.071]	113	0.052 [0.045–0.067]	97	0.055 [0.046–0.066]	98	0.32	0.29	0.73	0.75
Triglycerides in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	98	0.6	0.89	0.84	0.57
Triglycerides in very large VLDL	mmol/l	0 [0–0.0031]	113	0 [0–0]	97	0 [0–0.0048]	98	0.47	0.43	0.93	0.7
Triglycerides in large VLDL	mmol/l	0.011 [0–0.03]	113	0.0073 [0–0.023]	97	0.0037 [0–0.03]	98	0.42	0.69	0.42	0.86
Triglycerides in medium VLDL	mmol/l	0.043 [0.02–0.075]	113	0.032 [0.01–0.066]	97	0.032 [0.00046–0.069]	98	0.27	0.38	0.32	0.97
Triglycerides in small VLDL	mmol/l	0.055 [0.042–0.073]	113	0.047 [0.038–0.069]	97	0.05 [0.039–0.069]	98	0.17	0.16	0.42	0.83
Triglycerides in very small VLDL	mmol/l	0.039 [0.033–0.047]	113	0.035 [0.031–0.045]	97	0.037 [0.032–0.043]	98	0.078	0.084	0.24	0.81
Triglycerides in HDL	mmol/l	0.063 [0.055–0.071]	113	0.059 [0.051–0.067]	97	0.059 [0.052–0.065]	98	0.098	0.18	0.14	1
Triglycerides in large LDL	mmol/l	0.058 [0.051–0.068]	113	0.055 [0.048–0.064]	97	0.056 [0.048–0.06]	98	0.19	0.34	0.2	1
Triglycerides in medium LDL	mmol/l	0.017 [0.014–0.02]	113	0.016 [0.014–0.018]	97	0.016 [0.013–0.018]	98	0.18	0.31	0.2	1
Triglycerides in small LDL	mmol/l	0.0067 [0.0057–0.0081]	113	0.0061 [0.0053–0.0073]	97	0.0062 [0.0056–0.0073]	98	0.16	0.18	0.3	0.95
Triglycerides in very large HDL	mmol/l	0.0033 [0.0027–0.0041]	113	0.0033 [0.0025–0.0041]	97	0.0031 [0.0027–0.0039]	98	0.34	0.54	0.33	0.99
Triglycerides in large HDL	mmol/l	0.012 [0.0089–0.015]	113	0.012 [0.0078–0.015]	97	0.011 [0.0092–0.015]	98	0.7	0.76	0.74	0.98
Triglycerides in medium HDL	mmol/l	0.018 [0.014–0.022]	113	0.016 [0.013–0.021]	97	0.017 [0.013–0.022]	98	0.34	0.31	0.87	0.65
Triglycerides in small HDL	mmol/l	0.024 [0.021–0.029]	113	0.022 [0.019–0.028]	97	0.023 [0.02–0.029]	98	0.16	0.15	0.74	0.67
Cholesterol											
Total cholesterol	mmol/l	2 [1.6–2.5]	113	1.9 [1.7–2.3]	97	1.9 [1.6–2.3]	98	0.4	0.76	0.38	0.78
Total cholesterol minus HDL-C	mmol/l	1.2 [0.89–1.6]	113	1.1 [0.91–1.4]	97	1.1 [0.84–1.3]	98	0.2	0.52	0.19	0.73
Remnant cholesterol (non-HDL, non-LDL, -cholesterol)	mmol/l	0.62 [0.54–0.77]	113	0.59 [0.5–0.69]	97	0.58 [0.51–0.68]	98	0.046	0.11	0.07	0.99
VLDL cholesterol	mmol/l	0.25 [0.18–0.32]	113	0.22 [0.16–0.28]	97	0.21 [0.16–0.28]	98	0.039	0.082	0.07	0.99
Clinical LDL cholesterol	mmol/l	0.63 [0.32–1]	113	0.59 [0.38–0.85]	97	0.54 [0.27–0.83]	98	0.31	0.75	0.29	0.69
LDL cholesterol	mmol/l	0.57 [0.32–0.82]	113	0.58 [0.38–0.71]	97	0.54 [0.27–0.69]	98	0.29	0.72	0.28	0.63
HDL cholesterol	mmol/l	0.81 [0.73–0.95]	113	0.82 [0.72–0.95]	97	0.82 [0.74–0.92]	98	0.86	0.85	0.97	0.94
Cholesterol in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	98	0.63	0.88	0.87	0.6
Cholesterol in very large VLDL	mmol/l	0 [0–0.012]	113	0 [0–0]	97	0 [0–0.0097]	98	0.36	0.33	0.82	0.68
Cholesterol in large VLDL	mmol/l	0.013 [0–0.029]	113	0.011 [0–0.026]	97	0.0069 [0–0.026]	98	0.23	0.49	0.24	0.8
Cholesterol in medium VLDL	mmol/l	0.019 [0.0034–0.051]	113	0.016 [0.0025–0.035]	97	0.012 [0.00046–0.033]	98	0.14	0.37	0.14	0.87
Cholesterol in small VLDL	mmol/l	0.065 [0.045–0.086]	113	0.055 [0.039–0.075]	97	0.053 [0.036–0.074]	98	0.049	0.11	0.078	0.93
Cholesterol in very small VLDL	mmol/l	0.13 [0.11–0.14]	113	0.12 [0.11–0.14]	97	0.12 [0.11–0.13]	98	0.11	0.32	0.11	0.88
Cholesterol in IDL	mmol/l	0.38 [0.34–0.44]	113	0.36 [0.33–0.41]	97	0.36 [0.33–0.41]	98	0.18	0.31	0.21	0.99
Cholesterol in large LDL	mmol/l	0.37 [0.17–0.52]	113	0.35 [0.24–0.46]	97	0.33 [0.15–0.45]	98	0.28	0.7	0.27	0.68
Cholesterol in medium LDL	mmol/l	0.13 [0.076–0.18]	113	0.11 [0.075–0.16]	97	0.11 [0.061–0.16]	98	0.26	0.59	0.26	0.73
Cholesterol in small LDL	mmol/l	0.09 [0.072–0.11]	113	0.086 [0.072–0.1]	97	0.084 [0.067–0.098]	98	0.22	0.57	0.22	0.67
Cholesterol in very large HDL	mmol/l	0.1 [0.087–0.12]	113	0.11 [0.091–0.12]	97	0.1 [0.085–0.12]	98	0.63	0.99	0.73	0.64
Cholesterol in large HDL	mmol/l	0.23 [0.16–0.3]	113	0.23 [0.17–0.3]	97	0.23 [0.17–0.28]	98	0.77	0.93	0.93	0.75
Cholesterol in medium HDL	mmol/l	0.24 [0.2–0.28]	113	0.25 [0.21–0.29]	97	0.24 [0.21–0.28]	98	0.73	0.75	0.83	0.97
Cholesterol in small HDL	mmol/l	0.25 [0.23–0.28]	113	0.25 [0.22–0.28]	97	0.25 [0.22–0.28]	98	1	1	1	1
Total lipids											
Total lipids in lipoprotein particles	mmol/l	3.8 [3.2–4.6]	113	3.7 [3.2–4.3]	97	3.7 [3.1–4.2]	98	0.46	0.68	0.45	0.91
Total lipids in VLDL	mmol/l	0.55 [0.38–0.74]	113	0.47 [0.33–0.65]	97	0.45 [0.34–0.63]	98	0.09	0.14	0.15	1
Total lipids in LDL	mmol/l	0.89 [0.59–1.2]	113	0.87 [0.65–1.1]	97	0.84 [0.53–1]	98	0.26	0.68	0.25	0.66
Total lipids in HDL	mmol/l	1.7 [1.6–2]	113	1.8 [1.6–2]	97	1.8 [1.6–2]	98	0.86	0.87	0.92	0.98
Total lipids in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	98	0.6	0.88	0.84	0.58
Total lipids in very large VLDL	mmol/l	0 [0–0.022]	113	0 [0–0]	97	0 [0–0.02]	98	0.42	0.4	0.89	0.69
Total lipids in large VLDL	mmol/l	0.033 [0–0.074]	113	0.023 [0–0.065]	97	0.016 [0–0.069]	98	0.35	0.62	0.34	0.84
Total lipids in medium VLDL	mmol/l	0.093 [0.028–0.16]	113	0.071 [0.018–0.13]	97	0.055 [0.0013–0.12]	98	0.12	0.22	0.16	0.89
Total lipids in small VLDL	mmol/l	0.15 [0.11–0.2]	113	0.12 [0.097–0.18]	97	0.13 [0.093–0.18]	98	0.085	0.12	0.16	1
Total lipids in very small VLDL	mmol/l	0.24 [0.21–0.27]	113	0.23 [0.2–0.26]	97	0.23 [0.2–0.25]	98	0.078	0.17	0.099	0.98
Total lipids in IDL	mmol/l	0.57 [0.51–0.69]	113	0.56 [0.5–0.63]	97	0.55 [0.5–0.62]	98	0.23	0.4	0.24	0.96
Total lipids in large LDL	mmol/l	0.57 [0.32–0.75]	113	0.54 [0.39–0.68]	97	0.52 [0.3–0.66]	98	0.28	0.71	0.26	0.68
Total lipids in medium LDL	mmol/l	0.2 [0.13–0.27]	113	0.18 [0.13–0.24]	97	0.17 [0.11–0.24]	98	0.25	0.56	0.25	0.74
Total lipids in small LDL	mmol/l	0.15 [0.13–0.18]	113	0.15 [0.13–0.17]	97	0.14 [0.12–0.16]	98	0.17	0.48	0.16	0.7
Total lipids in very large HDL	mmol/l	0.21 [0.17–0.24]	113	0.21 [0.17–0.25]	97	0.2 [0.16–0.24]	98	0.69	1	0.77	0.69
Total lipids in large HDL	mmol/l	0.46 [0.34–0.58]	113	0.47 [0.35–0.59]	97	0.46 [0.37–0.56]	98	0.88	0.94	0.99	0.87
Total lipids in medium HDL	mmol/l	0.49 [0.42–0.57]	113	0.5 [0.44–0.57]	97	0.5 [0.44–0.56]	98	0.8	0.85	0.83	1
Total lipids in small HDL	mmol/l	0.61 [0.55–0.68]	113	0.6 [0.56–0.67]	97	0.6 [0.55–0.69]	98	0.95	0.94	0.98	1
Lipoprotein particle concentrations											
Total concentration of lipoprotein particles	mmol/l	0.011 [0.0094–0.013]	113	0.011 [0.0099–0.013]	97	0.011 [0.0097–0.013]	98	0.95	0.98	1	0.92
Concentration of VLDL particles	mmol/l	7.2e-05 [5.6e-05–9.4e-05]	113	6.4e-05 [4.9e-05–8e-05]	97	6.4e-05 [5e-05–8.1e-05]	98	0.087	0.15	0.14	1
Concentration of LDL particles	mmol/l	0.00034 [0.00023–5e-04]	113	0.00032 [0.00022–0.00043]	97	3e-04 [0.00019–0.00041]	98	0.15	0.46	0.15	0.69
Concentration of HDL particles	mmol/l	0.011 [0.009–0.012]	113	0.011 [0.0095–0.012]	97	0.011 [0.0093–0.012]	98	0.91	0.93	0.98	0.93
Concentration of chylomicrons and extremely large VLDL particles	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	98	0.61	0.87	0.85	0.59
Concentration of very large VLDL particles	mmol/l	0 [0–8.4e-07]	113	0 [0–0]	97	0 [0–7.3e-07]	98	0.41	0.37	0.86	0.71
Concentration of large VLDL particles	mmol/l	1.9e-06 [0–3.6e-06]	113	1.5e-06 [0–3.2e-06]	97	1.4e-06 [0–3.3e-06]	98	0.34	0.6	0.33	0.85
Concentration of medium VLDL particles	mmol/l	7.4e-06 [2e-06–1.3e-05]	113	5.3e-06 [1e-06–9.7e-06]	97	4.4e-06 [4.7e-08–9.6e-06]	98	0.092	0.2	0.12	0.88
Concentration of small VLDL particles	mmol/l	1.9e-05 [1.4e-05–2.6e-05]	113	1.5e-05 [1.2e-05–2.4e-05]	97	1.7e-05 [1.2e-05–2.4e-05]	98	0.11	0.14	0.21	0.99
Concentration of very small VLDL particles	mmol/l	4.1e-05 [3.7e-05–4.9e-05]	113	4e-05 [3.4e-05–4.6e-05]	97	3.9e-05 [3.5e-05–4.4e-05]	98	0.12	0.27	0.13	0.92
Concentration of IDL particles	mmol/l	0.00013 [9.3e-05–0.00017]	113	0.00012 [9.9e-05–0.00015]	97	0.00012 [9.4e-05–0.00015]	98	0.35	0.71	0.33	0.77
Concentration of large LDL particles	mmol/l	0.00016 [9.3e-05–0.00027]	113	0.00014 [9.2e-05–0.00021]	97	0.00013 [6.8e-05–0.00021]	98	0.15	0.47	0.15	0.68
Concentration of medium LDL particles	mmol/l	8.4e-05 [5.1e-05–0.00012]	113	7.1e-05 [4.5e-05–0.00011]	97	6.7e-05 [3.9e-05–0.00011]	98	0.18	0.41	0.2	0.76
Concentration of small LDL particles	mmol/l	1e-04 [8.6e-05–0.00012]	113	9.7e-05 [8.6e-05–0.00011]	97	9.5e-05 [8.3e-05–0.00011]	98	0.15	0.46	0.13	0.77
Concentration of very large HDL particles	mmol/l	0.00033 [0.00027–4e-04]	113	0.00033 [0.00027–4e-04]	97	0.00032 [0.00027–0.00038]	98	0.66	0.99	0.69	0.73
Concentration of large HDL particles	mmol/l	0.0013 [0.00087–0.0017]	113	0.0013 [0.00091–0.0018]	97	0.0013 [0.00097–0.0016]	98	0.88	0.96	0.97	0.87

Concentration of medium HDL particles	mmol/l	0.0023 [0.0018–0.0028]	113	0.0023 [0.0019–0.0028]	97	0.0023 [0.0019–0.0027]	98	0.86	0.88	0.91	0.99
Concentration of small HDL particles	mmol/l	0.0068 [0.0059–0.0076]	113	0.0068 [0.006–0.0075]	97	0.0066 [0.0059–0.0079]	98	0.95	1	0.97	0.94
Phospholipids											
Total phospholipids in lipoprotein particles	mmol/l	1.4 [1.2–1.6]	113	1.4 [1.2–1.6]	97	1.4 [1.2–1.6]	98	0.81	0.92	0.81	0.96
Phospholipids in VLDL	mmol/l	0.13 [0.096–0.19]	113	0.12 [0.083–0.16]	97	0.12 [0.082–0.16]	98	0.078	0.12	0.13	1
Phospholipids in LDL	mmol/l	0.24 [0.19–0.3]	113	0.23 [0.2–0.27]	97	0.23 [0.19–0.27]	98	0.31	0.67	0.3	0.72
Phospholipids in HDL	mmol/l	0.89 [0.78–0.99]	113	0.9 [0.8–1]	97	0.91 [0.81–0.99]	98	0.79	0.86	0.8	1
Phosphoglycerides	mmol/l	1.3 [1.1–1.5]	112	1.2 [1–1.4]	97	1.2 [1–1.4]	97	0.37	0.55	0.37	0.98
Ratio of triglycerides to phosphoglycerides	ratio	0.27 [0.24–0.36]	112	0.27 [0.23–0.32]	97	0.27 [0.23–0.33]	97	0.23	0.21	0.62	0.72
Total cholinines	mmol/l	1.5 [1.2–1.7]	113	1.4 [1.2–1.7]	97	1.4 [1.2–1.6]	98	0.4	0.58	0.41	0.97
Phosphatidylcholines	mmol/l	1.2 [1–1.4]	113	1.2 [1–1.4]	97	1.2 [1–1.4]	98	0.45	0.49	0.57	1
Sphingomyelins	mmol/l	0.19 [0.15–0.24]	113	0.19 [0.16–0.23]	97	0.18 [0.15–0.22]	98	0.51	0.98	0.67	0.5
Phospholipids in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	98	0.61	0.88	0.85	0.59
Phospholipids in very large VLDL	mmol/l	0 [0–0.0068]	113	0 [0–0]	97	0 [0–0.0045]	98	0.42	0.38	0.84	0.75
Phospholipids in large VLDL	mmol/l	0.0081 [0–0.018]	113	0.0068 [0–0.016]	97	0.0049 [0–0.016]	98	0.32	0.6	0.32	0.82
Phospholipids in medium VLDL	mmol/l	0.02 [0.0036–0.033]	113	0.013 [0.00096–0.025]	97	0.012 [7.6e-06–0.026]	98	0.094	0.22	0.12	0.84
Phospholipids in small VLDL	mmol/l	0.031 [0.021–0.042]	113	0.025 [0.017–0.034]	97	0.025 [0.017–0.035]	98	0.08	0.16	0.12	0.96
Phospholipids in very small VLDL	mmol/l	0.07 [0.063–0.08]	113	0.067 [0.059–0.076]	97	0.067 [0.06–0.073]	98	0.07	0.15	0.095	1
Phospholipids in HDL	mmol/l	0.15 [0.12–0.17]	113	0.14 [0.12–0.16]	97	0.14 [0.12–0.16]	98	0.46	0.79	0.46	0.78
Phospholipids in large LDL	mmol/l	0.13 [0.1–0.17]	113	0.13 [0.1–0.16]	97	0.13 [0.1–0.15]	98	0.36	0.75	0.34	0.76
Phospholipids in medium LDL	mmol/l	0.052 [0.038–0.068]	113	0.048 [0.038–0.062]	97	0.047 [0.035–0.062]	98	0.27	0.62	0.26	0.77
Phospholipids in small LDL	mmol/l	0.056 [0.051–0.061]	113	0.055 [0.05–0.059]	97	0.054 [0.05–0.058]	98	0.12	0.45	0.12	0.66
Phospholipids in very large HDL	mmol/l	0.099 [0.081–0.12]	113	0.1 [0.081–0.12]	97	0.097 [0.076–0.12]	98	0.72	0.99	0.81	0.71
Phospholipids in large HDL	mmol/l	0.22 [0.17–0.27]	113	0.22 [0.17–0.28]	97	0.22 [0.18–0.26]	98	0.91	0.92	1	0.92
Phospholipids in medium HDL	mmol/l	0.23 [0.21–0.27]	113	0.24 [0.21–0.27]	97	0.24 [0.21–0.27]	98	0.73	0.82	0.74	1
Phospholipids in small HDL	mmol/l	0.33 [0.3–0.38]	113	0.33 [0.31–0.37]	97	0.33 [0.31–0.38]	98	0.99	0.98	1	1
Particle diameter											
Average diameter for VLDL particles	nm	35 [34–36]	113	35 [34–35]	97	34 [33–36]	98	0.19	0.26	0.28	1
Average diameter for LDL particles	nm	23 [22–23]	113	23 [22–23]	97	23 [22–23]	98	0.15	0.5	0.14	0.63
Average diameter for HDL particles	nm	9.8 [9.6–9.9]	113	9.8 [9.6–10]	97	9.8 [9.6–9.9]	98	0.86	0.94	0.98	0.84
Apolipoproteins											
Apolipoprotein B	g/l	0.28 [0.2–0.4]	113	0.26 [0.2–0.33]	97	0.24 [0.18–0.33]	98	0.15	0.43	0.14	0.73
Apolipoprotein A1	g/l	0.76 [0.66–0.86]	113	0.78 [0.68–0.87]	97	0.78 [0.69–0.86]	98	0.77	0.82	0.8	1
Ratio of apolipoprotein B to apolipoprotein A1	ratio	0.37 [0.27–0.47]	113	0.35 [0.26–0.42]	97	0.31 [0.24–0.41]	98	0.052	0.23	0.054	0.69
Fatty acids											
Total fatty acids	mmol/l	7.7 [6.7–8.7]	113	7.3 [6.6–8.2]	97	7.4 [6.7–8.1]	98	0.15	0.21	0.25	0.95
Degree of unsaturation	degree	0.83 [0.71–0.94]	113	0.79 [0.65–0.91]	97	0.79 [0.67–0.89]	98	0.17	0.25	0.24	1
Omega-3 fatty acids	mmol/l	0.28 [0.21–0.39]	113	0.26 [0.19–0.36]	97	0.28 [0.18–0.38]	98	0.17	0.18	0.35	0.97
Omega-6 fatty acids	mmol/l	2.9 [2.6–3.1]	113	2.8 [2.6–3]	97	2.8 [2.6–3]	98	0.18	0.21	0.32	0.99
Polysaturated fatty acids	mmol/l	3.2 [2.9–3.5]	113	3 [2.8–3.3]	97	3.1 [2.8–3.3]	98	0.094	0.12	0.2	0.93
Monounsaturated fatty acids	mmol/l	1.7 [1.4–2]	113	1.5 [1.3–1.8]	97	1.5 [1.4–1.8]	98	0.16	0.2	0.28	0.93
Saturated fatty acids	mmol/l	2.7 [2.4–3.1]	113	2.6 [2.4–3]	97	2.7 [2.4–2.9]	98	0.31	0.38	0.41	1
Linoleic acid	mmol/l	1.9 [1.6–2.2]	113	1.7 [1.6–2]	97	1.8 [1.6–2.1]	98	0.052	0.056	0.18	0.87
Docosahexaenoic acid	mmol/l	0.34 [0.28–0.41]	113	0.31 [0.26–0.36]	97	0.31 [0.28–0.36]	98	0.021	0.031	0.1	0.65
Fatty acid ratios											
Ratio of omega-3 fatty acids to total fatty acids	%	3.8 [3.1–4.9]	113	3.6 [2.8–4.6]	97	3.5 [2.6–4.7]	98	0.23	0.26	0.37	0.99
Ratio of omega-6 fatty acids to total fatty acids	%	38 [36–40]	113	38 [37–40]	97	39 [37–40]	98	0.51	0.57	0.59	0.99
Ratio of polysaturated fatty acids to total fatty acids	%	42 [41–44]	113	42 [41–43]	97	42 [41–43]	98	0.87	0.94	0.86	0.98
Ratio of monounsaturated fatty acids to total fatty acids	%	22 [20–23]	113	21 [20–23]	97	22 [20–23]	98	0.29	0.28	0.56	0.85
Ratio of saturated fatty acids to total fatty acids	%	36 [35–37]	113	36 [36–37]	97	36 [36–37]	98	0.025	0.027	0.22	0.43
Ratio of linoleic acid to total fatty acids	%	25 [23–27]	113	24 [23–26]	97	24 [23–26]	98	0.059	0.055	0.36	0.52
Ratio of docosahexaenoic acid to total fatty acids	%	4.4 [4–4.9]	113	4.2 [3.9–4.5]	97	4.3 [4.1–4.6]	98	0.059	0.057	0.43	0.41
Ratio of polysaturated fatty acids to monounsaturated fatty acids	ratio	1.9 [1.8–2.1]	113	2 [1.8–2.1]	97	2 [1.8–2.2]	98	0.41	0.41	0.61	0.96
Ratio of omega-6 fatty acids to omega-3 fatty acids	ratio	9.7 [7.5–13]	113	11 [8.2–15]	97	11 [7.7–15]	98	0.23	0.25	0.39	0.98
Amino acids											
Alanine	mmol/l	0.46 [0.4–0.53]	113	0.45 [0.4–0.52]	97	0.53 [0.45–0.59]	98	1.7e-05***	0.8	3e-04**	6.9e-05***
Glutamine	mmol/l	0.49 [0.41–0.56]	72	0.49 [0.44–0.56]	55	0.48 [0.41–0.58]	52	0.85	1	0.86	0.88
Glycine	mmol/l	0.25 [0.22–0.29]	113	0.24 [0.22–0.27]	97	0.25 [0.22–0.28]	98	0.79	0.78	0.93	0.93
Histidine	mmol/l	0.1 [0.093–0.11]	113	0.11 [0.097–0.11]	97	0.11 [0.097–0.12]	98	0.35	0.65	0.55	0.83
Total concentration of branched-chain amino acids (leucine + isoleucine + valine)	mmol/l	0.4 [0.36–0.45]	113	0.39 [0.35–0.43]	96	0.41 [0.36–0.45]	98	0.19	0.8	0.46	0.16
Isoleucine	mmol/l	0.06 [0.05–0.069]	113	0.058 [0.05–0.069]	97	0.061 [0.053–0.068]	98	0.52	0.96	0.63	0.55
Leucine	mmol/l	0.12 [0.1–0.13]	113	0.11 [0.1–0.13]	97	0.11 [0.1–0.13]	98	0.72	0.81	0.97	0.74
Valine	mmol/l	0.22 [0.2–0.25]	113	0.22 [0.2–0.24]	96	0.23 [0.2–0.26]	98	0.064	0.82	0.17	0.075
Phenylalanine	mmol/l	0.086 [0.079–0.093]	113	0.085 [0.078–0.093]	97	0.089 [0.08–0.098]	98	0.19	0.99	0.24	0.27
Tyrosine	mmol/l	0.061 [0.055–0.067]	113	0.065 [0.057–0.072]	97	0.063 [0.055–0.07]	98	0.17	0.15	0.79	0.46
Glucose metabolism											
Glucose	mmol/l	4.3 [3.4–5.4]	113	4 [3.4–5]	97	4.4 [3.4–5.4]	98	0.68	0.87	0.9	0.67
Lactate	mmol/l	5.8 [4.8–7.4]	113	5.8 [4.5–7.6]	97	6.1 [5–7.6]	98	0.57	0.99	0.61	0.64
Pyruvate	mmol/l	0.11 [0.08–0.14]	71	0.12 [0.086–0.15]	54	0.11 [0.09–0.15]	52	0.89	0.88	0.95	1
Citrate	mmol/l	0.031 [0.025–0.038]	113	0.033 [0.026–0.038]	97	0.032 [0.025–0.039]	98	0.51	0.5	0.87	0.77
Glycerol	mmol/l	0.12 [0.091–0.15]	103	0.12 [0.092–0.14]	92	0.11 [0.092–0.15]	95	0.95	0.94	0.97	1
3-Hydroxybutyrate	mmol/l	0.16 [0.078–0.29]	113	0.15 [0.08–0.26]	96	0.17 [0.075–0.31]	97	0.63	0.93	0.83	0.58
Acetate	mmol/l	0.037 [0.029–0.047]	113	0.037 [0.03–0.048]	97	0.039 [0.031–0.049]	98	0.34	0.9	0.3	0.64
Acetoacetate	mmol/l	0.0012 [0–0.0031]	113	0.0013 [0–0.0036]	97	0.00041 [0–0.0027]	98	0.59	0.9	0.79	0.58
Acetone	mmol/l	0.037 [0.023–0.052]	113	0.035 [0.021–0.05]	97	0.037 [0.023–0.06]	98	0.45	0.78	0.81	0.42
Inflammation											
Glycoprotein acetyls	mmol/l	0.5 [0.45–0.58]	113	0.47 [0.44–0.55]	97	0.5 [0.46–0.55]	98	0.17	0.2	0.84	0.31

Overall p-value is calculated using the Kruskal-Wallis test, and for between group comparisons using the Dwass-Steel test. IQR: interquartile range. * p < 0.01, ** p < 0.001, *** p < 0.0001

Supplementary table 2 – Comparison of cord serum metabolites between metformin, insulin, and diet treatment groups with the patients in the metformin group requiring additional insulin excluded

	Unit	Diet		Insulin		Metformin		p-values			
		median [IQR]	n	median [IQR]	n	median [IQR]	n	Overall	Diet vs insulin	Diet vs metformin	Insulin vs metformin
Triglycerides											
Total triglycerides	mmol/l	0.36 [0.28–0.46]	113	0.31 [0.26–0.44]	97	0.32 [0.25–0.44]	76	0.25	0.24	0.54	0.92
Triglycerides in VLDL	mmol/l	0.14 [0.1–0.22]	113	0.12 [0.086–0.21]	97	0.13 [0.082–0.22]	76	0.29	0.28	0.54	0.98
Triglycerides in LDL	mmol/l	0.083 [0.073–0.095]	113	0.077 [0.068–0.089]	97	0.079 [0.067–0.087]	76	0.12	0.17	0.22	0.99
Triglycerides in HDL	mmol/l	0.057 [0.048–0.071]	113	0.052 [0.045–0.067]	97	0.056 [0.046–0.066]	76	0.32	0.29	0.87	0.65
Triglycerides in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	76	0.54	0.89	0.77	0.52
Triglycerides in very large VLDL	mmol/l	0 [0–0.0031]	113	0 [0–0]	97	0 [0–0.0052]	76	0.44	0.43	0.99	0.6
Triglycerides in large VLDL	mmol/l	0.011 [0–0.03]	113	0.0073 [0–0.023]	97	0.003 [0–0.031]	76	0.46	0.69	0.47	0.86
Triglycerides in medium VLDL	mmol/l	0.043 [0.02–0.075]	113	0.032 [0.01–0.066]	97	0.032 [0–0.071]	76	0.3	0.38	0.41	0.98
Triglycerides in small VLDL	mmol/l	0.055 [0.042–0.073]	113	0.047 [0.038–0.069]	97	0.051 [0.039–0.07]	76	0.18	0.16	0.59	0.75
Triglycerides in very small VLDL	mmol/l	0.039 [0.033–0.047]	113	0.035 [0.031–0.045]	97	0.038 [0.032–0.043]	76	0.093	0.084	0.46	0.65
Triglycerides in HDL	mmol/l	0.063 [0.055–0.071]	113	0.059 [0.051–0.067]	97	0.059 [0.051–0.066]	76	0.13	0.18	0.23	0.99
Triglycerides in large LDL	mmol/l	0.058 [0.051–0.068]	113	0.055 [0.048–0.064]	97	0.056 [0.048–0.061]	76	0.23	0.34	0.28	1
Triglycerides in medium LDL	mmol/l	0.017 [0.014–0.02]	113	0.016 [0.014–0.018]	97	0.016 [0.013–0.018]	76	0.22	0.31	0.3	1
Triglycerides in small LDL	mmol/l	0.0067 [0.0057–0.0081]	113	0.0061 [0.0053–0.0073]	97	0.0063 [0.0056–0.0074]	76	0.19	0.18	0.46	0.91
Triglycerides in very large HDL	mmol/l	0.0033 [0.0027–0.0041]	113	0.0033 [0.0025–0.0041]	97	0.0031 [0.0026–0.0039]	76	0.32	0.54	0.31	0.96
Triglycerides in large HDL	mmol/l	0.012 [0.0089–0.015]	113	0.012 [0.0078–0.015]	97	0.011 [0.009–0.014]	76	0.65	0.76	0.66	1
Triglycerides in medium HDL	mmol/l	0.018 [0.014–0.022]	113	0.016 [0.013–0.021]	97	0.017 [0.013–0.022]	76	0.3	0.31	0.97	0.49
Triglycerides in small HDL	mmol/l	0.024 [0.021–0.029]	113	0.022 [0.019–0.028]	97	0.024 [0.02–0.029]	76	0.13	0.15	0.96	0.27
Cholesterol											
Total cholesterol	mmol/l	2 [1.6–2.5]	113	1.9 [1.7–2.3]	97	1.9 [1.5–2.3]	76	0.27	0.76	0.28	0.53
Total cholesterol minus HDL-C	mmol/l	1.2 [0.89–1.6]	113	1.1 [0.91–1.4]	97	1 [0.84–1.4]	76	0.15	0.52	0.15	0.53
Remnant cholesterol (non-HDL, non-LDL-cholesterol)	mmol/l	0.62 [0.54–0.77]	113	0.59 [0.5–0.69]	97	0.58 [0.51–0.68]	76	0.043	0.11	0.069	0.95
VLDL cholesterol	mmol/l	0.25 [0.18–0.32]	113	0.22 [0.16–0.28]	97	0.21 [0.16–0.29]	76	0.047	0.082	0.1	0.98
Clinical LDL cholesterol	mmol/l	0.63 [0.32–1]	113	0.59 [0.38–0.85]	97	0.49 [0.26–0.84]	76	0.21	0.75	0.21	0.46
LDL cholesterol	mmol/l	0.57 [0.32–0.82]	113	0.58 [0.38–0.71]	97	0.49 [0.26–0.68]	76	0.21	0.72	0.22	0.43
HDL cholesterol	mmol/l	0.81 [0.73–0.95]	113	0.82 [0.72–0.95]	97	0.82 [0.71–0.92]	76	0.82	0.85	1	0.86
Cholesterol in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	76	0.57	0.88	0.81	0.54
Cholesterol in very large VLDL	mmol/l	0 [0–0.012]	113	0 [0–0]	97	0 [0–0.011]	76	0.35	0.33	0.95	0.57
Cholesterol in large VLDL	mmol/l	0.013 [0–0.029]	113	0.011 [0–0.026]	97	0.0072 [0–0.027]	76	0.28	0.49	0.32	0.85
Cholesterol in medium VLDL	mmol/l	0.019 [0.0034–0.051]	113	0.016 [0.0027–0.035]	97	0.011 [0–0.032]	76	0.12	0.37	0.13	0.62
Cholesterol in small VLDL	mmol/l	0.065 [0.045–0.086]	113	0.055 [0.039–0.075]	97	0.054 [0.038–0.074]	76	0.069	0.11	0.14	0.97
Cholesterol in very small VLDL	mmol/l	0.13 [0.11–0.14]	113	0.12 [0.11–0.14]	97	0.12 [0.11–0.13]	76	0.11	0.32	0.11	0.84
Cholesterol in IDL	mmol/l	0.38 [0.34–0.44]	113	0.36 [0.33–0.41]	97	0.36 [0.33–0.4]	76	0.15	0.31	0.17	0.94
Cholesterol in large LDL	mmol/l	0.37 [0.17–0.52]	113	0.35 [0.24–0.46]	97	0.3 [0.14–0.44]	76	0.19	0.7	0.2	0.43
Cholesterol in medium LDL	mmol/l	0.13 [0.076–0.18]	113	0.11 [0.075–0.16]	97	0.1 [0.058–0.15]	76	0.2	0.59	0.2	0.59
Cholesterol in small LDL	mmol/l	0.09 [0.072–0.11]	113	0.086 [0.072–0.1]	97	0.082 [0.067–0.098]	76	0.16	0.57	0.17	0.5
Cholesterol in very large HDL	mmol/l	0.1 [0.087–0.12]	113	0.1 [0.091–0.12]	97	0.1 [0.085–0.12]	76	0.52	0.99	0.63	0.51
Cholesterol in large HDL	mmol/l	0.23 [0.16–0.3]	113	0.23 [0.17–0.3]	97	0.23 [0.17–0.27]	76	0.68	0.93	0.84	0.65
Cholesterol in medium HDL	mmol/l	0.24 [0.2–0.28]	113	0.25 [0.21–0.29]	97	0.24 [0.2–0.28]	76	0.76	0.75	0.95	0.9
Cholesterol in small HDL	mmol/l	0.25 [0.23–0.28]	113	0.25 [0.22–0.28]	97	0.25 [0.22–0.28]	76	0.99	1	0.99	0.99
Total lipids											
Total lipids in lipoprotein particles	mmol/l	3.8 [3.2–4.6]	113	3.7 [3.2–4.3]	97	3.5 [3.1–4.2]	76	0.33	0.68	0.33	0.72
Total lipids in VLDL	mmol/l	0.55 [0.38–0.74]	113	0.47 [0.33–0.65]	97	0.45 [0.34–0.65]	76	0.11	0.14	0.21	1
Total lipids in LDL	mmol/l	0.89 [0.59–1.2]	113	0.87 [0.65–1.1]	97	0.78 [0.53–1.1]	76	0.2	0.68	0.21	0.47
Total lipids in HDL	mmol/l	1.7 [1.6–2]	113	1.8 [1.6–2]	97	1.8 [1.5–2]	76	0.86	0.87	1	0.91
Total lipids in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	76	0.55	0.88	0.78	0.52
Total lipids in very large VLDL	mmol/l	0 [0–0.022]	113	0 [0–0]	97	0 [0–0.021]	76	0.4	0.4	0.98	0.59
Total lipids in large VLDL	mmol/l	0.033 [0–0.074]	113	0.023 [0–0.065]	97	0.015 [0–0.072]	76	0.4	0.62	0.42	0.87
Total lipids in medium VLDL	mmol/l	0.093 [0.028–0.16]	113	0.071 [0.018–0.13]	97	0.055 [0–0.12]	76	0.13	0.22	0.19	0.88
Total lipids in small VLDL	mmol/l	0.15 [0.11–0.2]	113	0.12 [0.097–0.18]	97	0.13 [0.093–0.18]	76	0.11	0.12	0.27	0.99
Total lipids in very small VLDL	mmol/l	0.24 [0.21–0.27]	113	0.23 [0.2–0.26]	97	0.23 [0.2–0.25]	76	0.09	0.17	0.13	0.98
Total lipids in IDL	mmol/l	0.57 [0.51–0.69]	113	0.56 [0.5–0.63]	97	0.54 [0.5–0.62]	76	0.17	0.4	0.18	0.83
Total lipids in large LDL	mmol/l	0.57 [0.32–0.75]	113	0.54 [0.39–0.68]	97	0.48 [0.3–0.65]	76	0.19	0.71	0.19	0.44
Total lipids in medium LDL	mmol/l	0.2 [0.13–0.27]	113	0.18 [0.13–0.24]	97	0.16 [0.11–0.24]	76	0.19	0.56	0.19	0.6
Total lipids in small LDL	mmol/l	0.15 [0.13–0.18]	113	0.15 [0.13–0.17]	97	0.14 [0.12–0.16]	76	0.13	0.48	0.13	0.53
Total lipids in very large HDL	mmol/l	0.21 [0.17–0.24]	113	0.21 [0.17–0.25]	97	0.2 [0.17–0.23]	76	0.58	1	0.69	0.56
Total lipids in large HDL	mmol/l	0.46 [0.34–0.58]	113	0.47 [0.35–0.59]	97	0.46 [0.37–0.54]	76	0.81	0.94	0.96	0.78
Total lipids in medium HDL	mmol/l	0.49 [0.42–0.57]	113	0.5 [0.44–0.57]	97	0.49 [0.44–0.57]	76	0.86	0.85	0.96	0.97
Total lipids in small HDL	mmol/l	0.61 [0.55–0.68]	113	0.6 [0.56–0.67]	97	0.61 [0.55–0.68]	76	0.95	0.94	1	0.98
Lipoprotein particle concentrations											
Total concentration of lipoprotein particles	mmol/l	0.011 [0.0094–0.013]	113	0.011 [0.0099–0.013]	97	0.011 [0.0096–0.013]	76	0.82	0.98	0.92	0.77
Concentration of VLDL particles	mmol/l	7.2e-05 [5.6e-05–9.4e-05]	113	6.4e-05 [4.9e-05–8e-05]	97	6.4e-05 [5e-05–8.1e-05]	76	0.11	0.15	0.23	1
Concentration of LDL particles	mmol/l	0.00034 [0.00023–5e-04]	113	0.00032 [0.00022–0.00043]	97	0.00028 [0.00019–0.00041]	76	0.13	0.46	0.13	0.57
Concentration of HDL particles	mmol/l	0.011 [0.009–0.012]	113	0.011 [0.0095–0.012]	97	0.01 [0.0093–0.012]	76	0.83	0.93	0.99	0.79
Concentration of chylomicrons and extremely large VLDL particles	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	76	0.55	0.87	0.79	0.53
Concentration of very large VLDL particles	mmol/l	0 [0–8.4e-07]	113	0 [0–0]	97	0 [0–7.9e-07]	76	0.39	0.37	0.96	0.61
Concentration of large VLDL particles	mmol/l	1.9e-06 [0–3.6e-06]	113	1.5e-06 [0–3.2e-06]	97	1.2e-06 [0–3.4e-06]	76	0.4	0.6	0.42	0.89
Concentration of medium VLDL particles	mmol/l	7.4e-06 [2e-06–1.3e-05]	113	5.3e-06 [1e-06–9.7e-06]	97	4.4e-06 [0–9.7e-06]	76	0.098	0.2	0.14	0.86
Concentration of small VLDL particles	mmol/l	1.9e-05 [1.4e-05–2.6e-05]	113	1.5e-05 [1.2e-05–2.4e-05]	97	1.7e-05 [1.3e-05–2.4e-05]	76	0.14	0.14	0.35	0.93
Concentration of very small VLDL particles	mmol/l	4.1e-05 [3.7e-05–4.9e-05]	113	4e-05 [3.4e-05–4.6e-05]	97	3.9e-05 [3.5e-05–4.4e-05]	76	0.13	0.27	0.16	0.88
Concentration of IDL particles	mmol/l	0.00013 [9.3e-05–0.00017]	113	0.00012 [9.9e-05–0.00015]	97	0.00011 [9.3e-05–0.00014]	76	0.24	0.71	0.23	0.57
Concentration of large LDL particles	mmol/l	0.00016 [9.3e-05–0.00027]	113	0.00014 [9.2e-05–0.00021]	97	0.00013 [6.6e-05–0.00024]	76	0.12	0.47	0.13	0.51
Concentration of medium LDL particles	mmol/l	8.4e-05 [5.1e-05–0.00012]	113	7.1e-05 [4.5e-05–0.00011]	97	6.6e-05 [3.9e-05–0.00011]	76	0.15	0.41	0.17	0.65
Concentration of small LDL particles	mmol/l	1e-04 [8.6e-05–0.00012]	113	9.7e-05 [8.6e-05–0.00011]	97	9.2e-05 [8.3e-05–0.00011]	76	0.12	0.46	0.11	0.6
Concentration of very large HDL particles	mmol/l	0.00033 [0.00027–4e-04]	113	0.00033 [0.00027–4e-04]	97	0.00032 [0.00027–0.00037]	76	0.56	0.99	0.59	0.63
Concentration of large HDL particles	mmol/l	0.0013 [0.00087–0.0017]	113	0.0013 [0.00091–0.0018]	97	0.0013 [0.00098–0.0016]	76	0.8	0.96	0.9	0.78
Concentration of medium HDL particles	mmol/l	0.0023 [0.0018–0.0028]	113	0.0023 [0.0019–0.0028]	97	0.0023 [0.0019–0.0027]	76	0.88	0.88	0.99	0.93

Concentration of small HDL particles	mmol/l	0.0068 [0.0059–0.0076]	113	0.0068 [0.006–0.0075]	97	0.0066 [0.0059–0.0078]	76	0.96	1	0.98	0.95
Phospholipids											
Total phospholipids in lipoprotein particles	mmol/l	1.4 [1.2–1.6]	113	1.4 [1.2–1.6]	97	1.3 [1.2–1.6]	76	0.63	0.92	0.62	0.8
Phospholipids in VLDL	mmol/l	0.13 [0.096–0.19]	113	0.12 [0.083–0.16]	97	0.12 [0.081–0.16]	76	0.097	0.12	0.21	1
Phospholipids in LDL	mmol/l	0.24 [0.19–0.3]	113	0.23 [0.2–0.27]	97	0.22 [0.18–0.27]	76	0.22	0.67	0.23	0.52
Phospholipids in HDL	mmol/l	0.89 [0.78–0.99]	113	0.9 [0.8–1]	97	0.91 [0.79–0.99]	76	0.86	0.86	0.96	0.95
Phosphoglycerides	mmol/l	1.3 [1.1–1.5]	112	1.2 [1–1.4]	97	1.2 [1–1.3]	75	0.28	0.55	0.27	0.86
Ratio of triglycerides to phosphoglycerides	ratio	0.27 [0.24–0.36]	112	0.27 [0.23–0.32]	97	0.27 [0.23–0.33]	75	0.23	0.21	0.79	0.6
Total cholines	mmol/l	1.5 [1.2–1.7]	113	1.4 [1.2–1.7]	97	1.4 [1.2–1.6]	76	0.32	0.58	0.32	0.86
Phosphatidylcholines	mmol/l	1.2 [1–1.4]	113	1.2 [1–1.4]	97	1.2 [1–1.4]	76	0.35	0.49	0.41	0.92
Sphingomyelins	mmol/l	0.19 [0.15–0.24]	113	0.19 [0.16–0.23]	97	0.17 [0.15–0.22]	76	0.32	0.98	0.48	0.29
Phospholipids in chylomicrons and extremely large VLDL	mmol/l	0 [0–0]	113	0 [0–0]	97	0 [0–0]	76	0.55	0.88	0.78	0.53
Phospholipids in very large VLDL	mmol/l	0 [0–0.0068]	113	0 [0–0]	97	0 [0–0.0049]	76	0.4	0.38	0.95	0.63
Phospholipids in large VLDL	mmol/l	0.0081 [0–0.018]	113	0.0068 [0–0.016]	97	0.0044 [0–0.016]	76	0.38	0.6	0.4	0.87
Phospholipids in medium VLDL	mmol/l	0.02 [0.0036–0.033]	113	0.013 [0.00096–0.025]	97	0.012 [0–0.026]	76	0.094	0.22	0.13	0.8
Phospholipids in small VLDL	mmol/l	0.031 [0.021–0.042]	113	0.025 [0.017–0.034]	97	0.025 [0.017–0.035]	76	0.1	0.16	0.19	0.99
Phospholipids in very small VLDL	mmol/l	0.07 [0.063–0.08]	113	0.067 [0.059–0.076]	97	0.067 [0.06–0.074]	76	0.093	0.15	0.16	1
Phospholipids in LDL	mmol/l	0.15 [0.12–0.17]	113	0.14 [0.12–0.16]	97	0.13 [0.11–0.16]	76	0.34	0.79	0.35	0.58
Phospholipids in large LDL	mmol/l	0.13 [0.1–0.17]	113	0.13 [0.1–0.16]	97	0.12 [0.099–0.15]	76	0.24	0.75	0.23	0.52
Phospholipids in medium LDL	mmol/l	0.052 [0.038–0.068]	113	0.048 [0.038–0.062]	97	0.044 [0.034–0.062]	76	0.21	0.62	0.2	0.63
Phospholipids in small LDL	mmol/l	0.056 [0.051–0.061]	113	0.055 [0.05–0.059]	97	0.053 [0.05–0.058]	76	0.094	0.45	0.092	0.49
Phospholipids in very large HDL	mmol/l	0.099 [0.081–0.12]	113	0.1 [0.081–0.12]	97	0.096 [0.077–0.12]	76	0.61	0.99	0.73	0.58
Phospholipids in large HDL	mmol/l	0.22 [0.17–0.27]	113	0.22 [0.17–0.28]	97	0.22 [0.19–0.26]	76	0.86	0.92	0.99	0.85
Phospholipids in medium HDL	mmol/l	0.23 [0.21–0.27]	113	0.24 [0.21–0.27]	97	0.24 [0.21–0.27]	76	0.81	0.82	0.89	0.99
Phospholipids in small HDL	mmol/l	0.33 [0.3–0.38]	113	0.33 [0.31–0.37]	97	0.33 [0.31–0.37]	76	0.99	0.98	1	1
Particle diameter											
Average diameter for VLDL particles	nm	35 [34–36]	113	35 [34–35]	97	35 [33–36]	76	0.23	0.26	0.39	1
Average diameter for LDL particles	nm	23 [22–23]	113	23 [22–23]	97	23 [22–23]	76	0.13	0.5	0.13	0.52
Average diameter for HDL particles	nm	9.8 [9.6–9.9]	113	9.8 [9.6–10]	97	9.7 [9.6–9.9]	76	0.76	0.94	0.92	0.71
Apolipoproteins											
Apolipoprotein B	g/l	0.28 [0.2–0.4]	113	0.26 [0.2–0.33]	97	0.24 [0.18–0.33]	76	0.12	0.43	0.12	0.57
Apolipoprotein A1	g/l	0.76 [0.66–0.86]	113	0.78 [0.68–0.87]	97	0.78 [0.67–0.86]	76	0.83	0.82	0.95	0.97
Ratio of apolipoprotein B to apolipoprotein A1	ratio	0.37 [0.27–0.47]	113	0.35 [0.26–0.42]	97	0.31 [0.24–0.39]	76	0.061	0.23	0.072	0.67
Fatty acids											
Total fatty acids	mmol/l	7.7 [6.7–8.7]	113	7.3 [6.6–8.2]	97	7.3 [6.6–8.1]	76	0.15	0.21	0.26	0.99
Degree of unsaturation	degree	0.83 [0.71–0.94]	113	0.79 [0.65–0.91]	97	0.77 [0.66–0.9]	76	0.17	0.25	0.24	1
Omega-3 fatty acids	mmol/l	0.28 [0.21–0.39]	113	0.26 [0.19–0.36]	97	0.28 [0.18–0.39]	76	0.19	0.18	0.43	0.97
Omega-6 fatty acids	mmol/l	2.9 [2.6–3.1]	113	2.8 [2.6–3]	97	2.8 [2.6–3]	76	0.17	0.21	0.29	0.98
Polysaturated fatty acids	mmol/l	3.2 [2.9–3.5]	113	3 [2.8–3.3]	97	3.1 [2.8–3.4]	76	0.088	0.12	0.19	0.98
Monounsaturated fatty acids	mmol/l	1.7 [1.4–2]	113	1.5 [1.3–1.8]	97	1.5 [1.3–1.8]	76	0.17	0.2	0.34	0.94
Saturated fatty acids	mmol/l	2.7 [2.4–3.1]	113	2.6 [2.4–3]	97	2.6 [2.4–2.9]	76	0.28	0.38	0.35	0.99
Linoleic acid	mmol/l	1.9 [1.6–2.2]	113	1.7 [1.6–2]	97	1.7 [1.6–2]	76	0.053	0.056	0.2	0.97
Docosahexaenoic acid	mmol/l	0.34 [0.28–0.41]	113	0.31 [0.26–0.36]	97	0.31 [0.27–0.36]	76	0.023	0.031	0.12	0.75
Fatty acid ratios											
Ratio of omega-3 fatty acids to total fatty acids	%	3.8 [3.1–4.9]	113	3.6 [2.8–4.6]	97	3.5 [2.6–5]	76	0.24	0.26	0.42	1
Ratio of omega-6 fatty acids to total fatty acids	%	38 [36–40]	113	38 [37–40]	97	39 [37–41]	76	0.55	0.57	0.69	1
Ratio of polysaturated fatty acids to total fatty acids	%	42 [41–44]	113	42 [41–43]	97	42 [41–43]	76	0.88	0.94	0.88	0.97
Ratio of monounsaturated fatty acids to total fatty acids	%	22 [20–23]	113	21 [20–23]	97	22 [20–23]	76	0.31	0.28	0.72	0.78
Ratio of saturated fatty acids to total fatty acids	%	36 [35–37]	113	36 [36–37]	97	36 [36–37]	76	0.027	0.027	0.55	0.22
Ratio of linoleic acid to total fatty acids	%	25 [23–27]	113	24 [23–26]	97	24 [23–26]	76	0.064	0.055	0.49	0.52
Ratio of docosahexaenoic acid to total fatty acids	%	4.4 [4–4.9]	113	4.2 [3.9–4.5]	97	4.2 [4–4.7]	76	0.068	0.057	0.43	0.63
Ratio of polysaturated fatty acids to monounsaturated fatty acids	ratio	1.9 [1.8–2.1]	113	2 [1.8–2.1]	97	2 [1.8–2.1]	76	0.44	0.41	0.72	0.93
Ratio of omega-6 fatty acids to omega-3 fatty acids	ratio	9.7 [7.5–13]	113	11 [8.2–15]	97	11 [7.5–16]	76	0.25	0.25	0.48	0.98
Amino acids											
Alanine	mmol/l	0.46 [0.4–0.53]	113	0.45 [0.4–0.52]	97	0.53 [0.44–0.58]	76	0.00034**	0.8	0.0024*	0.00065**
Glutamine	mmol/l	0.49 [0.41–0.56]	72	0.49 [0.44–0.56]	55	0.48 [0.41–0.59]	41	0.96	1	0.95	0.97
Glycine	mmol/l	0.25 [0.22–0.29]	113	0.24 [0.22–0.27]	97	0.25 [0.22–0.28]	76	0.65	0.78	0.97	0.62
Histidine	mmol/l	0.1 [0.093–0.11]	113	0.11 [0.097–0.11]	97	0.11 [0.096–0.12]	76	0.46	0.65	0.49	0.91
Total concentration of branched-chain amino acids (leucine + isoleucine + valine)	mmol/l	0.4 [0.36–0.45]	113	0.39 [0.35–0.43]	96	0.4 [0.35–0.46]	76	0.34	0.8	0.64	0.31
Isoleucine	mmol/l	0.06 [0.05–0.069]	113	0.058 [0.05–0.069]	97	0.061 [0.053–0.068]	76	0.68	0.96	0.77	0.69
Leucine	mmol/l	0.12 [0.1–0.13]	113	0.11 [0.1–0.13]	97	0.11 [0.1–0.13]	76	0.79	0.81	1	0.86
Valine	mmol/l	0.22 [0.2–0.25]	113	0.22 [0.2–0.24]	96	0.23 [0.2–0.26]	76	0.2	0.82	0.37	0.2
Phenylalanine	mmol/l	0.086 [0.079–0.093]	113	0.083 [0.078–0.093]	97	0.088 [0.08–0.098]	76	0.29	0.99	0.33	0.37
Tyrosine	mmol/l	0.061 [0.055–0.067]	113	0.065 [0.057–0.072]	97	0.063 [0.054–0.069]	76	0.13	0.15	0.99	0.24
Glucose metabolism											
Glucose	mmol/l	4.3 [3.4–5.4]	113	4 [3.4–5]	97	4.3 [3.2–5.4]	76	0.84	0.87	0.99	0.86
Lactate	mmol/l	5.8 [4.8–7.4]	113	5.8 [4.5–7.6]	97	6.2 [5–7.6]	76	0.49	0.99	0.51	0.58
Pyruvate	mmol/l	0.11 [0.08–0.14]	71	0.12 [0.086–0.15]	54	0.11 [0.083–0.14]	41	0.87	0.88	0.99	0.91
Citrate	mmol/l	0.031 [0.025–0.038]	113	0.033 [0.026–0.038]	97	0.033 [0.026–0.039]	76	0.5	0.5	0.71	0.95
Glycerol	mmol/l	0.12 [0.091–0.15]	103	0.12 [0.092–0.14]	92	0.12 [0.092–0.15]	73	0.94	0.94	1	0.95
3-Hydroxybutyrate	mmol/l	0.16 [0.078–0.29]	113	0.15 [0.08–0.26]	96	0.15 [0.066–0.3]	75	0.87	0.93	0.99	0.85
Acetate	mmol/l	0.037 [0.029–0.047]	113	0.037 [0.03–0.048]	97	0.039 [0.031–0.05]	76	0.37	0.9	0.32	0.63
Acetoacetate	mmol/l	0.0012 [0–0.0031]	113	0.0013 [0–0.0036]	97	0.001 [0–0.0029]	76	0.9	0.9	1	0.95
Acetone	mmol/l	0.037 [0.023–0.052]	113	0.035 [0.021–0.05]	97	0.034 [0.023–0.061]	76	0.73	0.78	0.99	0.78
Inflammation											
Glycoprotein acetyls	mmol/l	0.5 [0.45–0.58]	113	0.47 [0.44–0.55]	97	0.5 [0.46–0.55]	76	0.19	0.2	0.89	0.37

Overall p-value is calculated using the Kruskal-Wallis test, and for between group comparisons using the Dwass-Stein test. IQR: interquartile range. * p < 0.01, ** p < 0.001, *** p < 0.0001